

BIONETICS

lethal assay-Contract FDA 71-268 & Compound FDA 71-40 (Dilauryl Thiodipropion Acid) 3/7/73 (Revised: 11/1/73)

ry of mutagenicity screening studies, heat-meditated assay cytogenetics domin

HOST-MEDIATED ASSAY CYTOGENETICS DOMINANT LETHAL ASSAY

FDA 71-40 lauryl Thiodipropionic Acid CONTRACT FUA /1-268

20014

SUMMARY OF MUTAGENICITY SCREENING STUDIES

HOST-MEDIATED ASSAY
CYTOGENETICS
DOMINANT LETHAL ASSAY
FDA 71-40
Dilauryl Thiodipropionic Acid
Contract FDA 71-268

SUBMITTED TO

Food & Drug Administration
Department of Health, Education and Welfare
Rockville, Maryland

SUBMITTED BY

Litton Bionetics, Inc. 7315 Wisconsin Avenue Bethesda, Maryland March 7, 1973

(Revised November 1, 1973)



Mr. Leonard Appleby, Contracting Officer Department of Health, Education & Welfare Public Heatlh Service Food & Drug Administration CA-212 5600 Fishers Lane, Room 5C-13 Rockville, Maryland 20852

Reference: Contract FDA 71-268

Dear Mr. Appleby:

Enclosed is the report of the mutagenicity screening studies conducted on FDA compound 71-40 Dilauryl Thiodipropionic Acid under the above referenced contract.

Included in this report are the results and raw data of the three tests conducted, the Host Mediated Assay, the Cytogenetic Studies and the Dominant Lethal Assay. Three(3) copies are being submitted for your review.

If there are any questions concerning this submission, or if additional information is required, please do not hesitate to contact me.

Sincerely yours,

LITTON BIONETICS, INC.

David P. A. Fabrizio

Principal Investigator

DPAF/vy Enclosure



TABLE OF CONTENTS

				Page	No.	
Ι.	REP	ORT	1			
	Α.	<u>In</u>	troduction	1		
	В.	<u>0b</u> ,	<u>jective</u>	4		
	С.	Con	npound	4		
		1.	Test Material	4		
		2.	Dosages	4		
	D.	Met	Methods			
	Ε.	Sum	Summary			
	F.	Res	ults and Discussion	7		
		1.	Toxicity	7		
			a. <u>In</u> <u>Vivo</u>	7		
			b. In Vitro	7		
			c. Toxicity Data Sheet	9		
		2.	Host-Mediated Assay	10		
			a. Host-Mediated Assay Summary Sheets	13		
			b. Host-Mediated Assay Data Sheets	18		
		3.	Cytogenetics			
		٠.	•	51		
			a. Cytogenetics Summary Sheets	52		
		4.	Dominant Lethal	56		
			a. Dominant Lethal Summary Tables	59		



TABLE OF CONTENTS (continued)

			Page	No			
II.	APP	APPENDICES (MATERIAL AND METHODS)					
	Α.	A. Animal Husbandry B. Dosage Determination					
	В.						
	С.	Mutagenicity Testing Protocols	78				
		1. Host Mediated Assay	78				
		2. Cytogenetic Studies	82				
		3. Dominant Lethal Assay	86				
	D.	Supplementary Materials and Methods	88				
		1. Host Mediated Assay <u>In Vitro</u> and Formulae	88				
		2. Cytogenetics $\underline{\text{In}}$ $\underline{\text{Vitro}}$ Preparation of Anaphase Chromosomes	91				
		3. Dominant Lethal Statistical Analyses	92				
	Ε.	References					
	F.	Abbreviations - Symbols and Explanations 97					
	G.	Dominant Lethal					
		Statistical Analyses Computer Print-Out Sheets					
		(Submitted Separately)					



I. REPORT

A. Introduction

Litton Bionetics, Inc. has investigated the possible mutagenicity of compounds selected and provided by the Food & Drug Administration under Contract 71-268. LBI's investigation utilized the three mammalian test systems herein described--Host-Mediated Assay, Cytogenetic studies and Dominant Lethal Assay. These tests provide information as to the types of genetic damage caused by environmental compounds -- pesticides, chemicals, food additives, drugs, and cosmetics.

The Host-Mediated Assay is based upon the assumption that the action of a mutagen on the genetics of bacteria is similar to that in man. This is further strengthened by the use of an eukaryotic organism (e.g., Saccharomyces cerevisiae). Since the mutation frequencies are well established for the indicator organism, any deviation due to the action of the test compound is readily detectable. As some compounds are mutagenic in bacteria and not in the host animal, and vice versa, this test is able to differentiate an action which may have been due to hosts' ability to detoxify or potentiate a suspected mutagen. This action is dependent upon the ability of the compound to gain access to the peritoneal cavity. Coupled with the direct action of the compound on the indicator organism in vitro, the assay provides a clear insight into host mediation of mutagenicity.

Cytogenetics provides a valuable tool for the direct observation of chromosomal damage in somatic cells. Alteration of the chromosome number and/or form in somatic cells may be an index of mutation. These studies utilized examination of bone marrow cells arrested in C-metaphase from rats treated with test compound as compared to positive and negative control animals. If muta-



tional changes occur the types of damage expected due to the action of chemicals are structural rearrangements, breaks and other forms of damage to the chromosomal complement of the cells exposed.

For the <u>in vitro</u> cytogenetic studies, we have a more rapid and inexpensive means of determining chromosomal damage. This is accomplished by observing cells in anaphase. As the chromatids separate and move along the spindle, aberrations may occur. Chromatids which do not migrate to the daughter cells may lead to uneven distribution of parts or of entire chromatids (mitotic nondysjunction). These give rise to "side arm" bridges which have been interpreted as point stickiness or localized failures of chromosome duplication point errors. These aberrations (bridges, pseudochiasmata, multipolar cells, acentric fragments, etc.) are extremely sensitive indicators of genetic damage.

The Dominant Lethal Test is an accurate and sensitive measure of the amount and type of fetal wastage which may occur following administration of a potential mutagen. Dominant lethal mutations are indicators of lethal genetic lesions. The effects of mutagens on the chromosomal complement of the spermatozoa of treated males results in alterations of form and number of chromosomes. Structural rearrangements and aneuploidy may lead to the production of non-viable zygotes, early and late fetal deaths, abortions



and congenital malformations. In addition, aberrations could lead to sterility or reduced reproductive capacity of the F_1 generation. The action of a mutagen on specific portions of spermatogenesis is also apparent in this test.



B. Objective

The purpose of these studies is to determine any mutagenic effect of the test compound by employing the Host Mediated Assay, Cytogenetics Studies and the Dominant Lethal Assay, both <u>in vivo</u> and <u>in vitro</u> tests are employed with the cytogenetic and microbial test systems. These tests and their descriptions are referenced in the Appendices A through F.

C. Compound

1. Test Material

Compound FDA 71-40, Dilauryl thiodipropionic acid 10-769, as supplied by the Food and Drug Administration.

2. Dosages

The animals employed, the determination of the dosage levels and the route of administration are contained in the technical discussions.

The dosage levels employed for compound FDA 71-40 are as follows for Cytogenetics Studies in vivo in rats.

Low Level	50	mg/kg
Intermediate Level	500	mg/kg
LD5	5000	mg/kg
Negative Control	salir	ie .
Positive Control (TEM*)	0.3	mg/kg

The dosage levels employed for compound FDA 71-40 are as follows for Host Mediated Assay in vivo in mice.

Low Level	50	mg/kg
Intermediate Level	500	mg/kg
LD ₅	5000	mg/kg
Negative Control	salin	
Positive Control		
(EMS **)	350	mg/kg
(DMN***)	100	mg/kg

- * Triethylene Melamine
- ** Ethyl Methane Sulfonate
- *** Dimethyl Nitrosamine



The dosage levels employed for compound FDA 71-40 are as follows for the Dominant Lethal Assay in vivo in rats.

Low Level	50	mg/kg
Intermediate Level	500	mg/kg
LD ₅	5000	mg/kg
Negative Control	salin	e
Positive Control (TEM*)	0.3	mg/kg

The <u>in vitro</u> cytogenetics studies were performed employing three logarithmic dose levels.

Low Level	5.0 mcg/ml
Medium Level	50.0 mcg/ml
High Level	500.0 mcg/ml
Negative Control	· saline
Positive Control (TEM*)	0.1 mcg/ml

The discussion of this test is contained in the technical discussions.

D. Methods

The protocols employed are explained in Appendices C and D.

E. Summary

1. Host Mediated Assay

Compound FDA 71-40 produced no significant reversion or recombinant increases in <u>Salmonella</u> strain TA-1530 or <u>Saccharomyces</u> strain D-3 respectively.

The results from tests using <u>Salmonella</u> strain G-46 indicated that this compound induced reversion in both the acute and subacute trials. A slight dose response was observed in the acute trials but not in the subacute trials.

Repeat tests of the subacute trials indicated the compound induced reversion, although the results were dose independent.

All in vitro tests were negative.



2. Cytogenetics

- (a) <u>In vivo</u> The compound produced no detectable significant aberration of the bone marrow metaphase chromosomes of rats when administered orally at the dosage levels employed in this study.
- (b) <u>In vitro</u> The compound produced no significant aberration in the anaphase chromosomes of human tissue culture cells when tested at the dosage levels employed in this study.
 - 3. This compound was considered to be non-mutagenic in rats in the dominant lethal assay when using the dosages employed in this study.

F-RESULTS AND DISCUSSION

1. TOXICITY

a. <u>In vivo</u>

Two male rats with an average body weight of 370 grams were given compound FDA 71-40 on an acute basis of 5000 mg/kg of body weight. The compound was in a suspension of 0.85% sterile saline and was administered by gastric intubation. All animals appeared normal during treatment and for an additional five days post-treatment observation. Necropsies of these animals on day six revealed no gross morphological change in the organs examined. The work was repeated with a group of ten male albino rats with an average body weight of 383 grams with the same findings. In the experiment 5000 mg/kg was administered at the high level, 500 mg/kg at the intermediate level, and 50 mg/kg at the low level. These dosages were employed in both the acute and the subacute in vivo studies. Animals in the acute studies were given a single dose of the compound. The subacute study animals were given the same dosages as those in the acute study, each day for five consecutive days 24 hours apart.

b. In vitro

The compound was suspended in 0.85% saline at the concentrations listed. It was introduced into culture tubes containing WI-38 cells in a logarithmic phase of growth. The cells were observed for cytopathic effect (CPE) and the presence of mitoses at 24 and 48 hours.



Tube No.	No. of cells	Conc. mcq/ml	<u>CPE</u>	<u>Mitosis</u>
1	5×10 ⁵	1000	+	+
2	ıı.	1000	+	+
3	н	500	-	+
4	II.	500	-	+
5	H	100	-	+
6		100	-	+
7	11	50	-	+
8	11	50	-	+
9	tt	10	•••	+
10	ii	10	-	+

Since a CPE was observed at 1000 mcg/ml a closer range of concentrations was employed, as follows.

1	1x10 ⁵	1000	+	+
2	H	1000	-	+
3	11	7 50	<u>+</u>	+
4	ıı	750	-	+
5		500	-	+
6	il	5 00	-	+
7	11	250	-	+
8	n	250	-	+
9	11	125	-	+
10	II	125	_	+

The high level used was 500 mcg/ml. The intermediate level used was 50 mcg/ml and the low level 5 mcg/ml.



c. TOXICITY DATA

COMPOUND FDA 71-40

This compound was administered at an extremely high concentration σf 5000 mg/kg with no abnormal effect observed in the animals.

Solvent: 0.85% sterile saline

Animals: Two (2) male rats--average weight 370 grams (observation

period six [6] days).

Ten (10) rats--average weight 383 grams (observation period

five [5] days).

Range finding:

Dose	No. Dead/No. Animals	Necropsy and Day of Death
5000 mg/kg	0/2	None
5000 mg/kg	0/10	None

There were no abnormal gross pathology findings in the animals dosed at 5000 mg/kg and a determination of an $\ensuremath{\text{LD}_{50}}$ was not performed.

2. Host Mediated Assay

Compound FDA 71-40 produced no significant reversion or recombinant increases in <u>Salmonella</u> strain TA-1530 or <u>Saccharomyces</u> strain D-3 respectively.

The results from tests using <u>Salmonella</u> strain G-46 indicated that this compound induced reversion in both the acute and subacute trials. A slight dose response was observed in the acute trials but not in the subacute trials.

Repeat tests of the subacute trials indicated the compound induced reversion, although the results were dose independent.

All in vitro tests were negative.



Compound: 71-40 Dilauryl Thiodipropionic Acid

		In Vivo			
Indicator Strain	In Vitro	Possible Low Recoveries	Controls Other Comments		
TA-1530 11/3/72 NO + Acutes 11/1/12 PC, NO Subscrites	pos.	NC PC AL AI AH SANC SAL SAI SAH	NC OK 1. The recoveries of some of the subacute trials were low and slightly elevated the reversion freq. The results are still in line and should be acceptable		
G-45	pos.	NC PC AL AI AH SANC SAL SAI SAI	1. Acute doses are positive and show a dose response. PC A little low 2. Subacute doses are positive. No dose response. SANC OK response.		
D3 PC, NC 5/1/12 acutes 5/5/12 subuntes	pos.	NC PC AL AI AH SANC SAL SAI SAH	1. No problems NC OK PC OK SANC OK		

and my comments regarding the lower than expected positive control as compound 36 and my comments regarding the lower than expected positive control results apply to this compound. All of the other data should be acceptable. The fact that some of the subacute recoveries for TA-1530 were a little low was not reflected in the reversion frequencies which were well in line with what would be expected. G-46 results indicate mutagenicity we have by this compound. TA-1530 and D3 were negative.

Compound: FDA-71-40 (Repeat)

			n Vivo	
Indicator Strain	<u>In Vitro</u>	Possible Low Recoveries	Controls	Other Comments
TA-1530	pos.	NC PC	NC	
		AL	PC	
	neg.	AI		
N . +		AH	SANC	
Not repeated		SANC		
		SAL		
		SAI		
		SAH		
G-46				
u-40		NC	NC OK 1.	All three dose levels
	pos.	PC ·	No on 1.	appear positive and
Subacutes only	poor	AL .	PC OK	show a slight dose
	neg.	AI		response.
	_	AH	SANC OK	
		SANC		
		SAL		
		SAI		
		SAH		
D3				
•		NC	NC	
	pos.	PC		
		AL	PC	
Not repeated	neg.	AI	0440	
	•	AH	SANC	
		SANC		
		SAL SAI		
		SAH		
		57 (7)		

Summary: Compound 40 shows mutagenic activity in G-46 at the subacute level when compared to the positive and negative controls. A slight dose response is seen, although it is probably not significant. I feel that these results should be accepted.

12

HOST MEDIATED ASSAY SUMMARY SHEETS

CONTRACT FDA 71-268 COMPOUND FDA 71-40 DILAURYL THIODIPROPIONIC ACID



HOST MEDIATED ASSAY

SUMMARY SHEET

OUTLIERS RELOVED

COMPOUND: FD	A 71-40	SALMO	NELLA		SACCHAROM'	YCES D-3
	TA15	5C	G-46	6-46		
	MMF (X 10E-8)	MFT/MFC	MMF (X 10E-3)	MFT/MFC	MRF (X 101-5)	MRT/MRC
ACUTE						
Signature of the state of the s	.58 8.00	13.79	.45 9.5% 2.11	21.13	4.80 53,17 5,02	11.70
हरून, हिन्दे हिन्दे	1.84 .41 1.45	2.66 .71 2.50	2.21 4.01 5.36	4.69 10.02 11.91	5,19 6,85	1.05 1.71 1.43
SUBACUTE						
**************************************	.6 6 1.96	2.97	.45 5.79	12,37	4, 93 4,88	•54
54, 54	1,22 2,35	1,05 3,55	5,62 5,99	15,16 13,31	4,27 4,4 7	.67 .91
20	2,00	<i></i>	0,00	1000	7678	• > 1
IN VITRO	TA1530	G = 46	% CONC	D-3 % SURVIVAL	R X 108	5
NC				•		

PC

STOP

HOST MIDIATED ASSAY

SUM 'ARY SHEET

OUTLIERS INCLUCED

CV, PO. HE: FE		SALMO		_	SACCHAROMY	CES D-3
	TA15	3 ⁰	G-#	5		
	NKF (% 102-8)	MFT/MFC	MMF (x 102-3)	MFT/MFC	>>F (X 10E≈5)	MRTZMRC
ACOTE NO PL A A	.58 8.00 1.78 .81 1.71	13.7 9 2.75 2.85	\$ 52 \$ 52 \$ 52 \$ 53 \$ 55 \$ 55	17.61 3.00 8.00 9.33	4,50 55,73 5,73 5,22 7,86	11.87 1.00 1.71 1.64
STANCUTE NO SA SA SA	.35 2,35 1,98 2,35	2,03 2,27 2,85	.54 5.02 6.82 5.99	10,04 12,03 11,09	4.63 4.63 4.55 4.25	1.00 •98 •92
IN VI7⊴C TCPD	TA1530	6≈¥6 -	* CONC 10.0	D#3 % SURVIVAL 81.2	11	5
NC PC	+	- +	1.0	100.0 42. 7	5 381	

HOST MEDIATED ASSAY (REPEAT)

SUMMARY SHEET

OUTLIERS REMOVED

COMP	CUND:	FDA	71-40

		SALMONEL		_	SACCHAROMYCES D-3	
	TA15	TA1530)		
	MMF (X 10E-8)	MFT/MFC	MMF (X 10E-8)	MFT/MFC	MRF (X 10E-5)	MRT/MRC
ACUTE						
NC	1.00		.62		1.00	
PC	0.	0.	16.46	26.55	0.	0.
AL	0.	0.	0.	0.	0.	G.
ΑI	. 0.	0.	0.	0.	0.	0.
AH .	0.	0.	0.	0.	0.	0.
SUBACUTE						
NC	1.00		.62		1.00	
SL	0.	0.	5.62	9.06	0.	0.
SI	0.	0.	6.36	10.26	0.	0.
SH	0.	0.	6.08	9.81	0.	0.
IN VITRO	TA1530	G-46		D - 3		
			% CONC	% SURVIVAL	R X 10E	5
NC						

PC

STOP SRU'S:.6

!SWITCH IN\$:MC606

!DIV

HUST MEDIATED ASSAY (REPEAT)

SUMMARY SHEET

OUTLIERS INCLUDED

COMPOUND: FDA 71-40

	SALMON				SACCHAROMYCES D-3	
	TA153	30	G-46			_
	MMF (X 10E-8)	MFT/MFC	MMF (X 10E-8)	MFT/MFC	MRF (X 10E-5)	MRT/MRC
ACUTE						
NC	1.00		.62		1.00	
PC	0.	0.	16.46	26.55	0.	0.
AL	0.	0.	0.	0.	0.	Ō.
AI	0.	0.	0.	0.	0.	0.
AH	0.	0.	0.	0.	0.	0.
SUBACUTE						
NC	1.00		.62		1.00	
5 L	0.	0.	5.62	9.06	0.	0.
SI	0.	0.	6.03	9.73	0.	0.
5H	0.	0.	6.33	10.21	0.	0.
IN VITRO	TA1530	G-46		D-3		
	.,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	0-40	% CONC	% SURVIVAL	R X 10E	:5
ИС						
PC						

STOP PC SRU'S:.5 !OFF

USAGE ON 07/30/73 AT 10:25:23

SRU'S:3.8 ELAPSED TIME: 00:10:48

9

GOOD BYE.

HOST MEDIATED ASSAY DATA SHEETS

CONTRACT FDA 71-268 COMPOUND FDA 71-40 Dilauryl Thiodipropionic Acid



Host Mediated Assay - Adjusted Raw CFU x $10^7/0.6$ ml

Step 1: Technician set counter - plates on counter.

Step 2: Automatic equipment accumulates counts on 3 plates of 10^{-6} dilution as CFU x $10^{7}/0.6$ ml.

Step 3: Automatic equipment multiplies count obtained in step 1 by 0.1666666666667 to obtain total count/ml at 108.

Step 4: Automatic check of result of step 3. TC x $10^8 \div 0.1667 = CFU \times 10^7/0.6 \text{ ml}$

Step 5: Technician was to record the true raw CFU x $10^7/0.6$ ml in log book, however, through error the computer provided the Column B check figure as the raw count.

To clarify the problem Column A is headed Adjusted Raw CFU X $10E^7/0.6~\text{ml}$ in each case where the check figure was provided as the raw count.



COMPOUND: FDA 71-40 ORGANISM: SALMONELLA TA1530

DOSE LEVEL: MEGATIVE CONTROL - SALINE

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: NOVEMBER 3, 1972

	Α	В	C	D
1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-	OAN FILV	TOTAL CFU X	TOTAL HO. MUTAHTS X	MUTATION FRE (C/B)
COTHAL FORDER		1023/1.0ML	10E0/1.0ML	X 10E-9
1	31.70	5 • 88	3.00	•57
2 3	35.20	5.67	2.00	•34
	21.70	3.62	2.00	• 55
4	47.00	7.83	1.00	• 13
5	27.10	4.5%	$4 \bullet 60$	• 39
b	17.60	2.93	3.00	1.03
7	<u> 22 • 50</u>	√ 3• 75	00 • بے 5 • 00	• 5 % • 5 %
- မိ မှ	34 • 00	5.67		
9	43.80	7.53	5.00	•63
	IMALS EQUALS ITH ZERO MUTAH	TS EGUAL 1		
		COL. 8 (x 1018)	CoL. C (* 1026)	COL. D (x 103-8)
	MEAT	5.20	2.78	\$ 5 g
	RANGE	4.70	4.00	•90
	MAX	7.63	5.00	1.02
	MIL	2 • 93	1.00	.13
OUTLE				♥ #. ₩!

CO GOUND: FDA 71-40 ORGANISM: SALMONELLA TAISON

DUST LEVEL: MOSITIME CONTROL - DAN - 100 MOVEG

THEATMERT: IN VIVE. ORAL. ACUTE DATE STARTED: NOVEMBER 1, 1972

	Α	В	TOTAL NO.	D MUTATION
MOIMAL No DOUR	RAW CFU X 1067/0.08L	TOTAL CEU X 1668/1.0ML	MUTANTS X 10EU/1.0ML	FRE (CZB) X 10E-3
1	35 . 20	5.37	ქმ•00	6.45
4	22.30	3•3€	42.00	11.05
<u>3</u>	31.70	5 • 20	23.00	5.30
11	19∙20	3.20	აე•00	9.37
5	28.40	4.75	22.00	4.65
ťı	31 • 00	5.17	ავ∙00	6 • 97
7	23.10	3• 65	47.00	12.21

NO. OF CONTANTED EQUALS 1
TOTAL OFF OUT OF ANGE EQUALS 2

	COL. U	COL. C	COL. D
	(X 10, 8)	(X 10E0)	(X 105-€)
₩EA. i	4.05	34.71	$6 \cdot 60$
RANGE	2.67	25.00	7.56
$\mathbf{x}_{\mathbf{A}^{\prime}\mathbf{A}}$	5.67	47.00	12.21
61h	3.20	22.00	4.65

NO OUTLIERS

C 5085F 02 DEC 72 15:18:51 USER CFU007 200

SECONDS OF THE OFFICE OF THE SECONDS

COMPOUND: FDA 71-40 ORGANISM: SALMONELLA TA153)

(USE LEVEL: LOW - 50 MU/KG

TREADMENT: IN VIVO, ORAL, ACUTE DATE STARTED: NOVEMBER 3, 1972

HIMAL Limerik	A RAW (FU X 1087/0.0%L	B TOTAL CFU X 10:8/1.0ML	C TOTAL NO. MUTANTS X 1080/1.UML	D MUTATION FRE (C/B) X 10E-9
1	25.50	4.25	μ•00	• 99
2	56.40	6.07	5•00	• 94
i,	11.60	1.9:	2.00	1.83
	73.40	12.23	3.00	.25
5	16.70	2.70	7•00	2.51
6	8.40	1.40	5•00	3.57
7	16.20	2.70	5 • 0 0	1.85
B	20.80	3.47	4 • 0 0	1.15
	IMALS EQUALS OUT OF HANGE E	d LQUALS 2		
		COL • B (X 100B)	CuL. C (X 10E0)	COL. • D (X 105-8)
	≥EAR	4.15	4.50	1.55
	RANGE	10.:3	5.00	3.33
	MAX	12.23 1.50	7.00 2.00	3.67 .25

NO OUTLIERS

COMPOUND: FDA 71-40 ORGANISM: SALMOMELLA TAISSO

DOSE LEVEL: INTERNEDIATE - 500 MG/KG

THE ATMENT: IN VIVO. ORAL. ACUTE DATE STARTED: NOVEMBER 3, 1972

	Α	8	C NO	D
N. HAAL	RAW CFU X	TOTAL CHU X	TOTAL NO. MUTANTS X	MUTATION FRE (CZP)
1076ER	1057/0.6ML	1000/1.0ML	1080/1.01L	X 10E-
1	36.30	6. 05	J+00	• Sn
2 3	21 • 1 ü	3 • 5 ≥	3•00	• 8%
ა	45 • 0 0	7. 50	2.00	• 27
4 5	20.30	3. •30	1.00	• 30
	35 ⊷ ⊍6	5.97	1.00	•17
5 7	ច្ចប្ត•ខិត្ត	16.15	2.00	• 200
8	37.00	6.17	4.00	•65
9	38•10 39•40	6+35 6+57	2•00 3•00	•31 •46
		9	3,00	• 1/ \
		Cot. B	CoL. C	COL. D
	15.	(X 10.8)	(X 10E0)	(X 102-8)
	AEA*	£ • 18	2.33	• 11
	RANGE	6.75	3.00	• 6 <u>9</u>
	MAX	10.13	4.00	• 55
O OUTLIER	MIN	3.08	1.00	• 1.7

COMPOUND: FDA 71-40 ORGANISM: SALMONELLA TA153%

DOSE LEVEL: HIGH - 5000 MOZKO

THEATMENT: IN VIVO. ORAL. ACUTE DATE STARTED: NOVEMBER 3, 1972

	Α	ដ	C Total No.	D MUTATION	
ANTIGAL	RAW CFU X	TOTAL CEU X	MUTANTS X	FRE (C/P)	
NUMBER	1057/0.6ML	1088/1.0ML	10E3/1.09L	X 10E-8	
1	18.10	3• 6€	4.00	1.33	
Ĺ	12.40	2.07	5.00	2.42	
ے خ	6.30	1.00	4.00	3.31	*
4.	10.10	1.08	2.00	1.10	
5	13.30	2.22	2.00	•99	
ti	7 • 00	1.17	1.00	• 35	
7	42.20	7 • 0.3	3• 00	655	
в	9•70	1.62	J•10	1.86	
9	21.80	3.60	£ • 60 °	2.20	
dia of all	IMALS EQUALS	9			
TOTAL CEU	OUT OF RANGE I				

	COL. B	COL. C	COL. D
	(X 1013)	(x 10E0)	(X 100-8)
MEAN	2.51	3.89	1.71
RANGE	5 • 93	7.00	2.96
MAX	7,03	(i • 0 i)	3.81
MIN	1.05	1.00	•35

* SUMMARY WITH OUTLIERS REMOVED

	COL. B (X 10:8)	COL. C (A 10E0)	COL. D (x 109+3)
ALK 1	$i_{\epsilon} \bullet \cdot \cdot 0$	3.33	1.00
RANUE	5.87	7.00	1.67
MAX	7.03	8.00	5.42
MIN	1.17	1.00	.85

ORGANISM: SALMONELLA TAIBBO COMPOUND: FUA 71-40

DUSE LEVEL: HEGATIVE CONTROL - SUPACUTE TRIALS

TELATMENT: IN VIVO. GRAL. ACUTE DATE STARTED: NOVEMBER 1. 197 /

ANIMAL Notalk	A RAW CEU X 1027/3.6/mg	B TOTAL CAU X 1028/1.0ML	C TOTAL NO. MUTAHTS X 1000/1.04L	D MUTATION FRE (CZB) X 108-9	
TOTAL CLU	8.70 16.40 17.89 16.80 12.30 13.40 16.20 22.70 IMALS EQUALS OUT OF KANGE NOTAN		3.00 2.00 1.00 1.00 2.00 2.00 4.00	2.07 .73 .34 .36 .47 .90 .79 1.06	*
	MEAN RANGE NAX MIN	COL+ B (X 103) 2+00 2+03 3+78 1+45	CoL. C (λ 10E9) 2.00 3.00 4.00 1.00	COt • 5 (X 10+6) •83 1•73 2•07 •34	

* SUMMARY WITH OUTLIERS REMOVED

	COL. A	CUL. C.	cot. D
	(8, 01, 8)	(X 10E0)	(X 105-8)
SEATI	2.016	1 • Ca	•66
RANGE	1.05	3.0 i	- 12
MAX	5.78	4 • 0 i	1.06
10114	2.43	1.09	• 4

COMPOUND: FDA 71-40 ORGANISM: SALMONELLA TA1530

DOSE LEVEL: LOW - 50 MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: NOVEMBER 1, 1972

ANIMAL NUMBER		8 TOTAL CFU X 10E8/1.0ML	C TOTAL NO. MUTANTS X 1000/1.0ML	D MUTATION FRE (CZ3) X 10E-8	
1 2	29•40 3•30	4.90 1.38	8.00 7.00	1.03 5.06	*
3 4 5	21.60 22.00 16.90	3+60 3+67	8.00 6.00	2.22 1.64	•
6 7 8	13.40 10.30	2.82 2.23 1.72	3.00 7.00 6.00	1.07 3.13 3.50	
No. of	10.70 AHIMALS EQUALS &		1.00	•50	
NO. OF	CONTAMINATED EQUALS	COL. B (X 10E6)	COL. C (X 10E0)	COL. D (X 10E-6)	
	MEAN RARGE MAX MIN	2.76 3.52 4.90 1.38	5.75 7.00 3.00	2.35 4.50 5.06	

* SUMMARY WITH OUTLIERS REMOVED

	col. n	col. c	COL. D
	(X 3.078)	(X 1080)	(X 100-8)
MERG	11,95	5.57	1.90
RAPSE	3 . 1 , 1,	7.00	2.93
19 A X	(1 🕳 🖟 .)	F.00	ວັ. ເນປ
MIN	1.72	1.00	•56

COMFOUND: FDA 71-80

ORGANISM: SALMOMELLA TAIDOR

DOSE LEVEL: INTERMEDIATE - SOD MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: NOVEMBER 1, 1972

AHIMAL MUSISER	A RAW CFU X 10E7/0.6ML	B TOTAL CFU X 1088/1.0ML	C TOTAL HO. MUTANTS X 10EO/1.OML	D MUTATION FRE (CZB) X 10E-8	
1 2 3 4 5 6 7 8	17.20 6.70 31.90 55.10 11.10 17.00 16.30 23.30	2.87 1.45 5.32 9.35 1.85 2.33 2.38	2.00 5.00 4.00 13.00 12.00 4.00 1.00 2.00	•70 3•45 •75 1•59 6•49 1•41 •56 •52	*
HO. OF CU	IMALS EDUALS NEAMINATED EOUA IEH ZERO MUYANT	.LS 1 'S EQUAL 1			
	MEATI RANGE MAX MIN	COL. B (X 10E8) 3.79 7.90 9.35 1.45	COL. C (X 1000) 5.33 12.00 13.00	COL. 0 (X 10, -3) 1.05 6.13 6.49	

* SUMMARY WITH OUTLIERS REMOVED

	COL. B	COL. C	COL. D
	(X 1058)	(X 10E0)	(X 106-8)
MEAN	4.07	4.43	1.22
RANGE	7.90	12.00	3.09
MAX	9.35	13.00	3.45
MIN	1.45	1.00	• 36

COMPOUND: FDA 71-40 ORGANISM: SALMONELLA TA1530

DOSE LEVEL: HIGH - 5000 MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: NOVEMBER 1, 1972

	A	в	С	D
			TOTAL NO.	MUTATION
ANIMAL	RAW CFU X	TOTAL CFU X	MUTANTS X	FRE (C/B)
NUMBER	10E7/J.6ML	1008/1.0ML	10E0/1.0ML	X 102-8
1	40.20	6.70	9,00	1.34
S	44.60	7.43	11.00	1.43
3	10.40	1.75	1.00	•58
4	47.10	7.85	5,00	• 04
5	42.80	7.13	6.00	• 84
6	18.00	3.00	19.00	6•ಎ3
7	41.10	6.65	16.00	2.34
ಕ	17.10	2.85	15.00	5.26
NO. OF AM	IMALS EQUALS	8		
NO. OF CO	NTAMINATED ECU	NLS 1		
SAMPLES W	ITH ZERO MUTAN	rs Equal 1		
		COL. B	COL. C	COL. D
		(X TOE8)	(X 10E0)	(X 10200)
	MERN	5.44	10.25	2.35
	RANGE	6.12	16.00	5.73
	MAX	7.65	1 0 • 0 0	6.33
	MIII	1.73	1.00	• ນຢ
				

NO OUTLIERS

COMPOUND: FDA 71-40 ORGANI M: SALMONELLA 0-46

WESE LEVEL: NEGATIVE CONTROL - SALINE

THEATHERT: IN VIVO, ORAL, ACUTE DATE STARTED: MAY 19, 1972

	A	В	С	С
AGISAL	ADJUSTED RAW CHU X	TOTAL CEU X	TOTAL NO. MUTANYS X	GUTATION
National R	101//0.68L	10E8/1.0ML	101.0/1.0ML	FRE (C/6) X 104-3
.1	200	4.71	1.00	• - 1
27	54.96	9.13	4.00	• 44
5	21.46	4.34	1.00	• 2 5
i.	43.14	6.59	5.00	•75
t,	34+(10	5.63	7.00	1.23
į, ,	37.38	6.25	4,00	≛ • ८ च • छाक्
7	20.62	3.37	1.00	• JU
<u>f.</u>	31.44	5.24	4.00	• 76
()	60.7c	10.12	3.00	• 50 • 50

TOTAL CHU OUT OF RAMSE EQUALS 1

	COL. H	CO: C	come o
No. 2	(X 10EF)	(X 1000)	(X) (- A)
Milnia Domini	○ • 1 4	. • 3.3	•
RA 19E	0.75	$\phi \bullet 00$	1.02
MAX	10.12	7 •00	1.025
MIL.	3.07	(• (i ()	• 4.4

* SUMBARY WITH GUTLIERS REMOVED

	COL. B	COL. C	COL. D
	(2 1 di 6)	(x (u_a)	(A 10a)
Mark	6.20	, • (1) 15	•45
KALL L	10 6 F	, • HO	•5:.
MAX	10.12	<.00 €	•76
MIV	3.37	1.00	•21

COMPOUND: FDA 71-40

_ UF

ORGANISMI SALMONELLA G-46

DUSE LEVEL: POSITIVE CONTROL - DMN - 100 MG/KG

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: MAY 19, 1972

arimal Number	A ADJUSTED RAW CRU X 10E7/0.6%L	B TOTAL CFU X Lowe/1.0ML	C TOTAL NO. MUTANTS X 10E0/1.0ML	D MUTATION FRE (C/D) X 105-8
1 25 45 5 7 8 9 0	11,82 29,68 23,94 22,68 33,78 33,90 20,13 39,20 37,70 32,55	1.97 4.93 3.99 5.73 5.65 3.35 6.23 5.43	20,00 34,00 38,00 24,00 44,00 75,00 23,00 97,00 61,00	10.15 6.23 9.52 6.35 7.22 13.27 6.26 14.65 9.71
NO. OF	ANIMALS EQUALS	10		
NO OUTL	MEAN RANGE MAX MIN	COL. B (X 10E8) 4.76 4.56 6.53 1.97	COL. C (X 10m0) 你的。90 77.60 97.00 20.00	COL. D (X 10£-4) 9,51 8.00 14.85 6.35

MUST PLUIATEL ASSAY REPORT DIEET

C - 17 July 1 D. 71-60 URGANISH: SALMONGELA 0-05 which LEVAL: Low - bu Merks THE ATMENTS OF VIVOR SHALL ACUTE DATE STARTLD: WAY 19, 1970 1 1) TO1 ... 1:0. **ADJUSTED** BUTATION A Dienst Har Cat W TOTAL CELL X NUTS ITS X FRE (0/0) Property 1027/000 L 1028/1.6gL 1000/1.031 3 105 - 3 25.32 ts . " " 4. . . 20.00 17. 10 1 . 13 2. . . 4.00 66. 16 3.70 3000 1. 35.0 5.90 5.00 • 47 30.000 · ** 12 5.8 2.30 ٠, 14. + 14 Owe Ca 9.00 1.59 7 34. 3 5.7 1.95 000 10.74 2.3 زن . . 00 2.1 . . 53.04 6.00 45.00 5.01 Mer AF Phillars Counties The face of the out of a large rounce of 1 Calle C Cina . 32 col. " (A 102 1) (X 10 6) (x 10"-0) L. 9 Himi 11.55 2.11 $\langle T_{ij} \rangle = 0$ 0.10 4.7 43.01 EA 30 P 3 S Ras U. 17.13 1. 2.0. 11 acres, i

HUST HUDIATED ALBAY REPORT EMEET

CO PUMBE: FDA /1-40

URGALISM: SALMONULLA 6-46

DOSE LEVEL: INTERMEDIATE - 500 MG/KG

TO EXTREMT: IN VIVE, UMAL, ACUTE

- UATE STARTED: MAY 19, 1972

Adlant NJack	ADJUSTED X 1007/0-05/L	B TOTAL COUX 10cHZ1•CHL	C TOTAL NO. NOTHERTS X 1025/1.00L	D MUTATION FRE (CZP) X 10E+2
i	27.54	4 (14	10.00	2 •86
2	20.64	3.44	9.00	2.62
ت	27.06	4.52	23.00	6.21
i ;	ພິລ•ີປ	4 • 1	2 % • 30	<u>ရှိ</u> ့်ချင်
/	214 • 15 to	4.11	15000	3.30
1)	32.55	5.45	J3 • 00	7.50
7	25.50	4.61	13.60	3.00

100. OF AMERICAN ECOALS 7
101. OF STAN ANIMALS ECOALS 1
TOTAL OF STANDE COURTS 2

	©©L • (3	COL. C	COL. D
	(x 10, 8)	(X 10E0)	(x 10°-a)
ME.A. r	is a wife	20•29	4.51
MARKET	1 • 49	29.00	$t_{r, \bullet}^{\epsilon} \cap \alpha$
A. A.	~ • # 3 3	ემ•0.	7.00
MIN	🛴 🗸 of 😘	9.00	2.42

170 OUTLIFE,5

Hast mulnific Abort Amport Solet

() 1 . (4 : FDA 71-4)

ORGANISH: SALMON LLA G-B.

DOUG LEVELS MICH - BUOD NO/KO

Total Started: MAY 10, 1074

nua A. L. Ulturalis	ADJUSTED HAR COU A ADJIZENTED ADJIZENTED	B TOTAL CHU X TUEOZI•OML	C TOTAL NO. MOTALITS X 10LL/1.00L	D AUTATION FRE (CAN) X 108-4
<u>.</u>	15.30	2.50	24.00	9.57
$\mathcal{C}_{\epsilon}^{i}$	11.10	1. • 6.60	S • 00	4.34
3	13.14	ن • 111	0 • 00	2.31
*** _{\$}	15.15	6 a 10 cm	7.00	2.77
,)	20.14	2.674	is a trul	1.
7	15.10	3.00	11.00	5.01
7	31 0	5021	26.00	5.57
*	1 hi	2 € 1.4	1: • 00	6.31
v 🍹	1.0 • ≥6	1011	17.00	9.34

I THE END OUT OF MALLE EQUALS 1

COL. B (X 1015)	CUL. C (x 1386)	COU. D (X 105+2)
G. • (1)	14.00	
å • . (i	_4 • U ↔	$\mathcal{E}_{\bullet} \circ \mathcal{E}$
$\omega \bullet \circ 1$. S • D €	9.4
1 + / 1	4 • U c	1.46
	(X 1000) 6 • (0 6 • (1	(X 1010) (X 1000) 0.00 14.00 0.00 24.00 0.00

HOST MEDIATED ASSAY REPORT SHEET

ComPodito: Fun 71-43

ORGANISM: SALMONELLA 6-40

DODE FEVEL: FOR - DO MONKO

THEATMENT: IN VIVE, ORAL, SUBACUTE DATE STARTED: MAY 19, 1972

A (18)(L N (18)(R	ADJUSTED RAW CEU X 1027/0.6ML	B TOTAL CFU X 10_8/1.03L	TOTAL NO. MUTANTS X MUJULOUL	D MUTATION FRE (C/R) X 10E-}
1 2 3 4 5 7 3 9	37.62 33.44 35.72 31.76 30.08 37.14 43.14 42.42 34.74 25.76	6.27 5.64 5.67 5.23 6.40 5.19 7.19 7.17 5.79	25.00 25.00 25.00 25.00 29.00 12.00 21.00 24.00	4.47 4.43 5.46 6.57 4.42 2.10 8.42 7.64
	FRALG EQUALS FLAN RANCE GAX MIN	COL • B (X 1968) 0 • 01 2 • 63 7 • 19 4 • • 6	24.00 COL. C (X 1980) U3.00 43.00 51.00	5•63 COL. 5 (X 105~8) 5.02 6.08 8.08

* SUMMARY WITH OUTLIERS REDIOVED

	COL. U	cor. c	cot n
	(x 10L8)	(X 10E0)	(x 101-8)
MEATE .	5.49	35.20	5.79
SANGE	ಪ್•≅3	37.00	4.005
PAA .	7.19	∂1•0 0	$\mathcal{B} \bullet \mathcal{B} \mathcal{B}$
514	4.4.6	24 · US	4.43

HELL PEDIATER AS AY REPORT SHEET

No reas : 100 71- 0

ORGINISM: SALMONILLEA G-40

EUSE LEVEL: INTERMEDIATE - 500 MOZKG

TALATAS T: 14 VIVO, ONAL, SUBACUTE DATE STARTED: MAY 19, 197

n al-dat Noonen	ADJUSTED FAM GEU X 1017/0.061	B TOTAL CHU X 1010/1•0ML	C TOTAL HO. MUTANTS X 1060/1.0ML	D MUTATION FRE (CZN) A 10"-
1 3 9 0 7 0 9	30.30 29.72 21.10 27.60 31.38 20.56 30.18 26.96 30.36	6.63 4.67 3.63 4.20 9.40 4.20 6.30 4.20	50.00 42.00 55.00 54.00 55.00 27.00 56.00 25.00	4.96 8.60 9.01 7.06 6.66 5.90 6.00 4.35
	INALS EGMALS AD ANIMALS EGUA		•	
NO OUTLAC	MENT MARGE PAA MIN	COL. (S (X 1808) 9.23 8.52 6.05 3.03	COL. C (X 10E0) 22.67 20.00 42.00 22.00	COL. D (X 108-8) 6.13 5.57 9.51 4.35

HUST MEDIATED ASSAY REPORT SHEET

COMPOURL: FDA 71-40

ORGANISM: SALMONELLA 6-46

bosh LEVEL: mich - 5000 Mg/KG

MU OUTLIFAS

TOLATECOTE 10 VIV. GRAL SUBACUTE DATE STARTED: MAY 19. 1972

	Α	В	C	D
	ADJUSTED		TOTAL NO.	MOTATION
Allking	RAW OFU X	TOTAL CFC X	MUTANTS X	FRE (C/H)
Michael	1007/3.0ML	1018/1.CHL	1000/1:0ML	X 10E-3
1 2	43.32	7. 82	21.00	2.91
	23.70	3.95	0 ل • د د د	6.35
j.	29.34	4.59	30.00	7.50
4į	20.52	4.4.	5 2•6 0	7.04
ŧ,	35.76	0.13	ئان•غد 2ۥ00	5.00
Ų	20∙20	4.50	ძე•მმ	6.39
7	34.32	5.72	21.00	3.67
Ö	27.78	4.00	64.00	5.13
9	24.48	4.0:	J5•00	9.31
	JATYAL, EDEVLA			
no• cr	DEAD ANIMALS COUNT	<u>د</u> 1		
		COL. B	COL. C	coL. n
		(X 10(B)	(X IDEO)	(X 10E-8)
	EAL	り・いち	28.67	5.09
	PA will	3. € 7	17.00	6.41
	AX	7.63	38.00	9.31
	MIN		9 1 . 0 9	S 11

COMPOUND: FDA 71-40 ORGANISM: SALMONELLA G-46

DOSE LEVEL: NEGATIVE CONTROL - SALINE

RANGE

MAX

MIN

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: JULY 9, 1973

ANIMAL NUMBER	A RAW CFU X 10E7/0.6ML	B TOTAL CFU X 10E8/1.0ML	C TOTAL NO. MUTANTS X 10EO/1.0ML	D MUTATION FRE (C/B) X 10E-8
1 3 4 5 6 7 8 9 10	53.90 54.80 35.70 50.30 41.30 44.20 83.70 61.90 52.10 96.30	8.98 9.13 5.95 8.38 6.88 7.37 13.95 10.32 8.68 16.05	2.00 5.00 2.00 6.00 4.00 6.00 8.00 7.00 7.00	.22 .55 .34 .72 .58 .81 .681 .93
NO. OF	ANIMALS EQUALS	10		
	ME AN	COL. B (X 10E8) 9.57	COL. C (X 10E0) 6.20	COL. D (X 10E-8) .62

10.10

16.05

5.95

13.00

15.00

2.00

.71

.93

.22

NO OUTLIERS

STOP SRU'S:.6

!switch ins:Mc662

!SAL

COMPOUND: FDA 71-40 ORGANISM: SALMONELLA G-46

DOSE LEVEL: POSITIVE CONTROL - DMN - 100 MG/KG

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: JULY 9, 1973

ANIMAL NUMBER	A RAW CFU X 10E7/0.6ML	B TOTAL CFU X 10E8/1.0ML	C TOTAL NO. MUTANTS X 10E0/1.0ML	D MUTATION FRE (C/B) X 10E-8
1 2 3 4 5 6 7 8 9 10	84.50 58.00 89.00 39.30 56.10 35.90 63.00 76.70 72.20 65.20	14.08 9.67 14.83 6.55 9.35 5.98 10.50 12.78 12.03	182.00 176.00 187.00 143.00 153.00 132.00 199.00 173.00 141.00	12.92 16.21 12.61 21.83 16.36 22.06 18.95 13.53 11.72 16.38
NO. OF AN	IMALS EQUALS	10		
	MEAN RANGE MAX MIN	COL. B (X 10E8) 10.67 8.85 14.83 5.98	COL. C (X 10E0) 166.40 67.00 199.00 132.00	COL. D (X 10E-8) 16.46 10.34 22.06 11.72

NO OUTLIERS

STOP SRU'S:.6 !SWITCH IN\$:MC663 !SAL

COMPOUND: FDA 71-40 ORGANISM: SALMONELLA G-46

DOSE LEVEL: LOW - 50 MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: JULY 9, 1973

	A	В	C TOTAL NO.	D MUTATION
ANIMAL NUMBER	RAW CFU X 10E7/0.6ML	TOTAL CFU X 10E8/1.0ML	MUTANTS X 10E0/1.0ML	FRE (C/B) X 10E-8
1	75.10	12.52	59.00	4.71
2	31.80	5.30	38.00	7.17
3	68.80	11.47	29.00	2.53
4	82.30	13.72	71.00	5.18
5	74.90	12.48	48.00	3.85
6	69.30	11.55	73.00	6.32
7	45.60	7.60	62.00	8.16
8	58.20	9.70	68.00	7.01

NO. OF ANIMALS EQUALS 8
NO. OF CONTAMINATED EQUALS 1
TOTAL CFU OUT OF RANGE EQUALS 1

	COL. B	COL. C	COL. D
	(X 10E8)	(X 10E0)	(X 10E-8)
MEAN	10.54	56.00	5.62
RANGE	8.42	44.00	5.63
MAX	13.72	73.00	8.16
MIN	5. 30	29.00	2.53

NO OUTLIERS

STOP SRU'S:.6

!SWITCH IN\$:MC664

!SAL

COMPOUND: FDA 71-40 ORGANISM: SALMONELLA G-46

DOSE LEVEL: INTERMEDIATE - 500 MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: JULY 9, 1973

ANI MAL NUMBER	A RAW CFU X 10E7/0.6ML	B TOTAL CFU X 10E8/1.0ML	C TOTAL NO. MUTANTS X 10E0/1.0ML	D MUTATION FRE (C/B) X 10E-8	
1 2 3 4 5 6 7 8 9	77.10 71.80 93.10 55.90 73.90 45.80 52.30 64.20 68.10	12.85 11.97 15.52 9.32 12.32 7.63 8.72 10.70 11.35	82.00 41.00 91.00 49.00 73.00 65.00 71.00 56.00 63.00	6.38 3.48 5.86 5.26 5.93 8.15 5.55	×

NO. OF ANIMALS EQUALS 9
NO. OF CONTAMINATED EQUALS 1

	COL. B	COL. C	COL. D
	(X 10E8)	(X 10E0)	(X 10E-8)
MEAN	11.15	65.67	6.03
RANGE	7.88	50.00	5.09
MAX	15.52	91.00	8.52
MIN	7.63	41.00	3.43

SUMMARY WITH OUTLIERS REMOVED

	COL. B	COL. C	COL. D
	(X 10E8)	(X 10E0)	(X 10E-8)
MEAN	11.05	68.75	6.36
RANGE	7.88	42.00	3.28
MAX	15.52	91.00	8.52
MIN	7.63	49.00	5.23

STOP SRU's:.7 !SWITCH IN\$:MC601 !DI

COMPOUND: FDA 71-40 ORGANISM: SALMONELLA G-46

DOSE LEVEL: HIGH - 5000 MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: JULY 9, 1973

ANIMAL NUMBER	A RAW CFU X 10E7/0.6ML	B TOTAL CFU X 10E8/1.0ML	C TOTAL NO. MUTANTS X 10E0/1.0ML	D MUTATION FRE (C/B) X 10E-8	
1 2 3 4 5 6 7 8 9	84.20 44.20 89.50 37.80 68.70 21.70 60.30 48.90 51.20 44.40	14.03 7.37 14.92 6.30 11.45 3.62 10.05 8.15 8.53 7.40	80.00 51.00 84.00 46.00 64.00 31.00 61.00 37.00 60.00 44.00	5.70 6.92 5.63 7.30 5.59 8.57 6.07 4.54 7.03 5.95	×

NO. OF ANIMALS EQUALS 10

	COL. B	COL. C	COL. D
	(X 10E8)	(X 10E0)	(X 10E-8)
MEAN	9.18	55.80	6.33
RANGE	11.30	53.00	4.03
MAX	14.92	84.00	8.57
MIN	3.62	31.00	4.54

* SUMMARY WITH OUTLIERS REMOVED

	COL. B	COL. C	COL. D
	(X 10E8)	(X 10E0)	(X 10E-8)
MEAN	9.80	58.56	6.08
RANGE	8.62	47.00	2.76
MAX	14.92	84 .0 0	7.30
MIN	6.30	37.00	4.54

STOP

SRU'S:.7 !SWITCH IDS:MC651 : PAPED PRINT ON SIX-PART PAPER

HOST MEDIATED ASSAY REPORT SHEET

COMPOUND: FDA 71-40 URGANISM: SACCHAROMYCES D-3

DOSE LEVEL: NEGATIVE CONTROL - SALINE

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: MAY 1, 1972

	A	В	C	D
Allidate	KAW CFU X	TOTAL CHU SCREENED X	TOTAL RECOMBINANTS	RECOMBICEU SCHEENED X
NUMBER	10E5/1.0ML	10E5/1.0> L	/1 • 0 mil.	10g-5
1	422.00	•140	3.00	7.11
2	173.00	•17	1.00	5.73
3	133.00	•13	• 0 0	• 0.0
Lj	648.00	• © \$	4.00	6.17
5	266+80	•27	2.00	7. 52
to	109.00	•11	1.00	9.17
7	1 98•00	•2,	• 00	• 00
8	371.00	• 37	1.00	2.70
y	181.00	•10	•00	•00
TOTAL		2.50	12.00	

NO. OF CONTAMINATED EQUALS 1

MEAN CIMEAN B # . 4.80

•		COL. B	COL. C	- col. D
		(X 405)	(人 10mg)	(X 10E+5)
	MEAN	• 28	1.33	4.27.
	RANGE	• ₹.· 4	4.00	9.17
	^{V}AX	• ()	4.0∂	9.17
	4114	• 1 1	• 0 U	• 6:0
1.6 25 Mars 2 12	,			

HE OUTLASHED

HUST MEDIATED ASSAY REPORT SHEET

COMPOUND: FDA 71-40 ORGANISM: SACCHAROMYCES D-3

DOSE LEVEL: POSITIVE CONTROL - EMS 350 MG/KG

TREATMENT: IN VIVO+ ORAL+ ACUTE DATE STARTED: MAY 1+ 1972

ANIMAL NUMBER	A RAW CFU X 10E5/1.0ML	B TOTAL CFU SCREENED X 10ED/1.UAL	C TOTAL RECUMBINANTS /1.0ML	D RECOMBZCFU SCREENED X 10E=5
1	414.00	•41	21.00	50.72
2	263.00	•2o	13.00	63.60
3	301.00	• ŠU	20.00	66.45
44	299.00	•30	14.00	46.82
5	232.00	.23	11.00	47.41
6	326.00	• 3 3	19.00	58.23
7	103.00	•10	00 ون	77.67
ઇ	266.00	.27	15.00	56 • 39
9	193.00	•19	12.00	62.18
1.0	392.00	•39	22.00	56.12
TOTAL		2.61	100.00	

NO. OF ANIMALS EQUALS 10

MEAN CYMEAN B = 35.95

	COL. B	COL. C	COL. D
	(X 1005)	(X 10E0)	(X 105~5)
MEAN	• ∉8	16.00	58.57
RAHGE	• .5 1	14.00	30.85
MAX	• 4 1	22.00	77.67
MIN	•10	8.00	46.82

* SUMMARY WITH OUTLIERS REMOVED

MEAN C/MEAN B = 56.17

	COL. B	col. c	col. n
	(x 10.5)	(x 10E0)	(X 103-5)
IL AN	• (1)	16.69	56.44
RANGE	\$2.	$11 \bullet 0 \oplus$	19.72
AA	• ~ 1	22.00	66 . "
MIN, COLOR	•19	11.0 J	46.82

HOST MEDIATED ASSAY REPORT SHEET

COMPOUND: FDA 71-40 ORGANISM: SACCHAROMYCES U-3

DOSE LEVEL: LOW - 50 MG/KG

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: MAY 1, 1972

	A	В	С	D
		TOTAL CFU	TOTAL	RECOMB/CFU
ANIMAL	RAW CFU X	SCREENED X	RECOMBINANTS	SCREENED X
NUMBER	10E5/1.0ML	10E5/1.0ML	/1.0ML	105-5
1	30 8 • 00	•31	2.00	6.49
2	290.00	•29	2.00	6.90
3	473.00	.47	3.00	6.34
4	421.00	•42	2.00	4.75
5	633 •00	•63	4.00	6.32
6	213.00	•22	1.00	4.59
7	423.00	• 42	1.00	2.36
8	287.00	• 29	1.00	3.48
9	531.00	•53	2.00	3.77
TOTAL		3. 58	18.00	
NO. OF AN	NIMALS EQUALS	9		
NO. OF DE	ALIOR PIEMTHA GAR	1 C 1		

NO. OF DEAD ANIMALS EQUALS 1

MEAN C/MEAN B = 5.02

		COL. B	COL. C	COL. D
		(X 10E5)	(X 10E0)	(X 10E-5)
	MEAN	•40	2.00	5.00
	RANGE	• 41	3.00	4.53
	MAX	•63	4.00	6.90
	MIN	•22	1.00	2.36
NO OUTLIERS				

44

HOST MEDIATED ASSAY REPORT SHEET

COMPOUND: FDA 71-40 ORGANISM: SACCHAROMYCES D-3

DOSE LEVEL: INTERMEDIATE - 500 MG/KG

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: MAY 1, 1972

ANIMAL NUMBER	A RAW CHU X 10ED/1.0ML	B TOTAL CAU SCREENED X 10E5/1.0ML	TOTAL RECOMBINANTS /1.UML	D RECOMB/CFU SCREENED X 10E-5
1 2 3 4 5	230.00 182.00 267.00 390.00 243.00 600.00 199.00	.23 .15 .25 .39 .24 .60 .20	2.00 1.00 3.00 3.00 3.00 7.00 1.00	8.70 5.89 10.45 7.69 12.35 11.67
8 TOTAL	433.00	2.55	1.00 21.00	2.31
	ANIMALS EQUALS DEAD ANIMALS EQUA		·	
MEAN C/	MEAN B =	5.19		
	1457.4.4	COL. 8 (X 10.5)	COL. C (X 10E0)	001.0 (X 100-5)

	COL. B	COL. C	001 0
	(X 10.5)	(X 10E0)	(X 10~~5)
MEAN	•32	2.63	7.96
RANGE	• 42	6.00	10.04
MAX	• 4.0	7.00	12.35
MIN	•18	1.00	2.31

NO OUTLIERS

HUST MEDIATED ASSAY REPORT MELT

CO (PUUNU: FDA 71-4) ORGANISM: SACCHARONYCES DE

DOSE LEVEL: HIGH - SOUD MG/KG

TREATMENT: IN VIVO, ORAL, ACUTE DATE STARTED: MAY 1, 1972

	A	B TOTAL CAU	C TOTAL	D RECOMPZCFU
AHIMAL	RAW CFU X	SCREENED X	RECOMBINANTS	SCREENED X
NUMBER	10E5/1.0ML	10E5/1.0-L	/1.UML	10E-5
1	521.00	• 5 d	7.00	13.44
\mathcal{Q}_{i}	5 07. 00	•51	4.00	7.89
خ خ	382.00	• 3 5	2.00	5.24
4	520.00	• 3 ≤	2.00	6.25
5	475.00	•47	3.00	6.39
Ćz	288.00	* k J	3.00	10.42
7	203.00	• 2 Q	1.00	4.93
ن	521.00	• 5.2	_ ц•∪0	7.65
9	222.00	· La da	1.00	4.50
TUTAL		3.44	27.00	

NO. OF ANIMALS EQUALS 9 TOTAL SCREENED OUT OF RANGE EQUALS 1

MEAN Ch LAN B = 7.83

	COL. B	COL. C	col. D
	(X 1025)	(X 10E0)	(X 10E-5)
MEAN	• 38	3.00	7.41
RANGE	• 52	6.00	3.93
MAX	•52	7.00	13.44
MIN	• d 0	1.00	4.50

* SUMMARY WITH OUTLIERS REMOVED

/ MEAN C/MEAN B = 6.80

	COLUMB	COL. • C	COL. D
	(2.30.5)	(x 1060)	(x 10-,→5)
loE A Se	 □ □ · C) 	2.000	6.66
RADGE	• 2	. 3.00	5.01
∂AX	•	4.00	10.00
MIN	• & 0	1.00	4.50

RA 6503.F 29 NOV 72 17:55:14 USER CFU307 200

HOST MEDIATED ASSAY REPORT SHEET

CUMPOUND: FDA 71-40 ORGANISM: SACCHAROMYCES D-3

DOSE LEVEL: NEGATIVE CONTROL - SUBACUTE TRIALS

TREATMENT: IN VIVO. ORAL. ACUTE DATE STARTED: MAY 5. 1972

	Α	8	C	.D
Y ! A	DESTRUCTION V	TOTAL CFU	TOTAL	RECOMB/CFU
AMIMAL	RAH CHU X	SCREENED X	RECOMBINANTS	SCREENED X
NUMBER	10ES/1.0ML	10世5/1.08世	/1.UML	10E-5
1	243.00	•24	1.00	4.12
2	428.00	•43	2.00	4.67
3	193.00	•19	1.00	5.13
ţţ	207.00	.21	1.00	4.83
5	351.00		2.00	5.70
6	333.00	.33	1.00	3.00
7	186.00	•19	.00	• 0.0
Ö	291.00	.29	2.00	6.87
9	411.00	•41	3.00	7.30
19	382.00	• 30	1.00	2.62
TOTAL		3.02	14.00	

NO. OF ANIMALS EQUALS 10

FIGAN CLEEN B = 4.63

	COL.∗ B	Col. C	col. D
	(X 10:5)	(X 10E0)	(X 105-5)
MEAN	• ∪ 0	1.40	4.43
RANGE	• 24	3.00	7.3n
MAX	•43	3.00	7.30
MIN	•19	• 0 0	•00

* SUMMARY WITH OUTLIERS REMOVED

MEAN C/MEAN B = 4.93

	COL. 9	Col. C	cot. • n
	(X 10.5)	(X 10E3)	(X 10~~5)
ERI	• 52	1.50	4. 2
KANGL	• 4.3	2.03	4.68
× AX	• 4 3	3.0	7. A
8:114	•19	1.0:1	2.62

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HOST MEDIATED ASSAY REPORT SHEET

COMPOUND: FDA 71-40 ORCANISM: SACCHAROMYCES D-3

DOSE LEVEL: LOW - 50 MG/KG

TREATMENT: IN VIVO, ORAL, SUBACUTE DATE STARTED: MAY 5, 1972

ARIMAL, NUMBER	A RAW CFU X 10E5/1.0ML	B TOTAL CHU SCREENED X 1025/1•02L	C TOTAL RECOMMINANTS /1.0ML	D RECOMPTOFU SCREENED X 102-5
1	125.00	•12	• 00	• 00
2	315.00	•31	1.00	3.17
2 3	189.00	• 1 y	ī•00	5.29
4	153.00	•15	• 0 0	• 0 0
5	368.00	•37	2.00	5.43
6	145.00	• 14	1.00	6.00
7	327.00	• 3.3	3.00	9.17
ũ	149.00	•10	1.00	6.71
9	382.00	• 35	Ε00	2.62
TOTAL		2.10	10.00	

NO. OF ANIMALS EQUALS 9
TOTAL SCREENED OUT OF RANGE EQUALS 1

MEAN CAMEAN B = 4.64

		COL· G	COL. C	col. • n
•		(% 10.5)	(~ 1000)	(x 100~5)
	MEAN	• = 4	1.1)	4.37
	RAHVE	• . 6	3.00	9.17
	TAX	• .	3 . U (i)	9.17
	AIII.	• # 2	• 0.1	• (-0)
. M. W. See	,			

NO CUTLICKS

HOST MEDIATED ASSAY REPORT SHELT

ORGANISM: SACCHAROMYCES D-COMPOUND: FDA 71-40

DOSE LEVEL: HIGH - 5000 MG/KG

THEATHENT: IN VIVO. ORAL. SUBACUTE DATE STARTED: MAY 5. 1972

ABIMAL NOMBLR	A RAW CFU X 10E5/1.0ML	B TOTAL CHU SCREENED X 10ES/1.00L	C TOTAL RECUMBINANTS /1.0al	D RECOMB/CFU SCREENED X 10E-5
1	207.00	.21	1.00	4.83
	422.00	• 44 c.	2.00	4.74
2 3	276.00	• & &	1.00	3 •62
4	413.00	• 4 1	2.00	4.64
Š	149.00	•15	•00	• 0.0
6	247.00	• 25	1.00	4.05
7	162.00	•10	1.00	6.17
ន់	412.00	•41	1.00	2.43
Ğ	208.00	•21	ī•00	4.81
10	563+00	∂ ₫•	3.00	5.33
TUTAL		3. 06	13.00	

NO. OF ANIMALS EQUALS 10

MEAN CAMEAN B = 4.25

MIN

	COL. 8 (X 10€5)	(X 10E0)	col. 0 (X 105-5)
MEAN	•.1	1.30	4.08
RANGE	• 41	3.00	6.17
MAX	• 56	3.00	6.17
MIN	• 4.5	•00	• 0 0

* SUMMARY WITH OUTLAERS REMOVED

MEAN CZMEAN B =	4.47		
	cot. B	COL. C	COL. D (X 100-5)
	(X 1015)	(x 10£0)	
bi€ Air	• ∪ <u>'</u>	1.44	4.54
MANAL.	• % ()	2.00	3.75
MAX	<u> ن پ</u> ن	გ.მა	6.17
MIN	• 16	1.00	2.43

3. Cytogenetics

In vivo

(1) Acute study

The chromosomal abnormalities observed in the positive controls were significantly higher than either the negative controls or the compound. The percentage of aberrations observed in the compound dosage groups was within the normal negative control values. The frequency of breaks in the negative controls was well within the range that we have seen in the past (0-6%). Mitotic indices were normal.

(2) Subacute study

The only aberrations observed in these groups were 3% breaks in the negative controls and 2% breaks in the high dose level.

These are within normal limits. Mitotic indices were normal.

In vitro

Anaphase preparations were examined in this test. The positive control compound produced a significantly higher percentage of aberrations on the chromosomes than the negative control or the test compound. Depression of the mitotic index due to the positive control was not as pronounced as in the <u>in vivo</u> test. The effect of the compound on the cells observed was negative. Negative controls were well within normal limits.

CYTOGENETICS

SUMMARY SHEETS

CONTRACT FDA 71-268 COMPOUND FDA 71-40 Dilauryl Thiodipropionic Acid



DILAURYL THIODIPROPIONIC ACID FDA 71-40 ACUTE STUDY METAPHASE SUMMARY SHEET

Compound	Dosage (mg/kg)	Time*	No. of <u>Animals</u>	No. of Cells	Mitotic*** Index %	% Cells with Breaks	% Cells with Reunion	% Cells Other Aber.**	% Cells with aber.
Negative Control	saline	6 24 48	3 3 3	_ 150 150 150	9 8 6	0 1 5	0 0 0	0 0 0	0 1 5
Low Level	50	6 24 48	5 5 5	250 250 260	12 10 10	0 0 0	0 0 0	0 0 0	0 0 0
Intermediate Level	500	6 24 48	5 5 5	250 250 250	13 7 8	0 3 0	0 0 0	0 0 0	0 3 0
High Level	500 0 -	6 24 4 8	· 5 5 5	250 250 250	12 13 13	0 1 2	0 0 0	0 0 . 0	0 1 2
Positive Control(TE	⁽⁴) 0.3	48	5	250	4	2 2	9	3(a)	31

^{*}Time of sacrifice after injection (hours)

**Cells that have polyploidy (P), sulverization (pp), or greater than 10 aberrations (a).

*** % of cells in mitosis:500 cells observed/animal.

DILAURYL THIODIPROPIONIC ACID FDA 71- 40 SUBACUTE STUDY METAPHASE SUMMARY SHEET

Compound	Dosage* (mg/kg)	No. of Animals	No. of Cells	Mitotic*** Index %	% Cells with Breaks	% Cells with Reunion	% Cells Other Aber.**	% Cells with aber.
Negative Control	saline	3	150	12	3	0	0	3
Low Level	50	5	250	13	0	0	0	0
Intermediate Level	500	5	250	13	0	0	0	0
High Level	5000	5	250	10	2	0	0	. 2

^{*}Dosage 1X/day X 5 days
**Cells that have polyploidy (P), pulverization (pp), or greater than 10 aberrations (a).
***% of cells in mitosis:500 cells observed/animal.

DILAURYL THIODIPROPIONIC ACID FDA 71-40

ANAPHASE SUMMARY SHEET

Compound	Dosage (mcg/ml)	. Mitotic** 	No. of Cells	% Cells with Acentric Frag.	% Cells with Bridges	% Multipolar Cells	% Cells Other Aber.*	% Cells with aber.
Low Level	5	2	100	0	0	0	0	0
Medium Level	50	1	100	0	0	0	0	0
High Level	500	5	100	0	0	0	0	0
Negative Control	saline	4	100	1	0	0	0	1
Positive Control (TEM)	0.1	2	100	13	6	0	0	19

^{*}Cells that have polyploidy (P), pulverization (pp), or greater than 10 aberrations (a).

**% of cells in mitosis:200 cells observed/dose level.

4. Dominant Lethal Assay

The interpretation of these data was made by Dr. David Brusick, Assistant Professor of Microbiology, Howard University, Washington, D.C., as a consultant to LBI.

a. Fertility Index

Acute Study - All the experimental groups were shown to differ significantly from the control group in week 7, having a lower fertility rate than expected.

Subacute Study - The low and high dose groups were shown to differ significantly from the control in week 3, with higher fertility rates than expected. Week 7 showed the same results as the acute study. Significant linear relationships with dose were shown for both of these weeks.

b. Average number of implantations per pregnant female

Acute Study - Significant differences in the experimental groups from the control group were seen in weeks 1, 3, 4, 6, and 7; decreases were shown in weeks 1, 3, 4, and 7. A significant relationship to dose was shown in weeks 1 and 3.

Subacute Study - A significant relationship with dose was shown in week 6, where the average implantations increased with dosage. The intermediate dose of week 7 showed a significant increase. The intermediate dose of week 2 was significantly decreased.

c. Average corpora lutea per pregnant female

Acute Study - A significant decrease in the intermediate dose group from the control group was shown in week 3. In week 4 the low and high dose groups showed significant decreases, and in the 8th week all experimental groups showed significant decreases. The results in all of these weeks were significantly related to dose.

Subacute Study - The intermediate dose group showed significant increases in weeks 6 and 7 and the high dose group showed a significant increase in week 6. In both weeks a significant relationship with dose was shown. Week 5 showed significant increases at the low and high doses which are dose related. The high dose of week 1 was significantly decreased.



d. Average preimplantation losses per pregnant female

Acute Study - Highly significant, dose related increases in the experimental groups were shown in weeks 1 and 7. Significant dose related decreases were shown in week 8.

Subacute Study - Significant, dose related increases in the intermediate and high dose groups over the control were shown in week 7.

The low dose group showed a significant increase over the control group in week 6. The high dose produced a significant decrease in week 1.

e. Average resorptions per pregnant female

Acute Study - The intermediate dose group in week 1 and the high dose in week 7 showed a significant increase over the control. In week 4 the intermediate and high dose groups showed significant dose related decreases from the control.

Subacute Study - Decreases at the high doses noted in weeks 1 and 2 were shown to have a significant relationship with dose. The low and high dose groups showed significant increases over the control group in week 3. In week 4 the intermediate dose group showed a significant decrease from the control.

f. Proportion of females with one or more dead implantations

Acute Study - The intermediate dose group showed significant increase in week 1. In week 4 both the intermediate and high dose groups showed significant dose related decreases.

Subacute Study - The high dose group differed significantly in weeks 1, 2, and 3, showing lower in weeks 1 and 2 and higher in week 3. The intermediate group was seen to have a significant decrease in week 4.



g. Proportion of females with two or more dead implantations Acute Study - Significant reduction at intermediate dose at week 3.

Subacute Study - In week 3 the high dose group showed a significant increase.

h. Dead implants / Total implants

Acute Study - Significant increases at the intermediate dose of week 1 and the high dose at week 7.

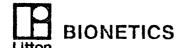
Subacute Study - Significant increases are observed for the low and high doses of week 3. Significant decreases were obtained at the high doses of weeks 1 and 2 and the intermediate dose of week 4.

DOMINANT LETHAL SUMMARY TABLES

CONTRACT FDA 71-268

COMPOUND FDA 71-40

DILAURYL THIODIPROPIONIC ACID



The Table

COYPOURD AR STUDY ACUTE

FFRTILITY INDEX

	ADITH DOSE	%	"ISTOFICAL CONTROL	NEGATIVE CONTROL	DOSE LEVEL 50.000 MG/KG	DOSE LEVEL 500.000 MG/KG	DOSP IPVEL 5000.000 MG/KG	POSITIVE CONTROL
	: !	1	109/159=0.69	13/20=0.65	12/20=0.60	13/19=0.69	9/20=0.45	10/13=0.56
		2	119/ 159 =0.75	16/19=0.85	15/20=0.75	16/19=0.85	13/18=0.73	11/19=0.58
			110/158=0.76	16/20=0.80	12/19=0.64	14/20=0.70	15/20=0.75	11/20=0.55
		4	1/6/160=0.85	18/20=0.90	18/20=0.90	19/20=0.95	17/20=0.85	16/20=0.80
! !		5	127/159=0.80	19/20=0.95	19/20=0.95	20/20=1.00	19/20=0.95	18/20=0.90
		į,	128/159=0.81	13/19=0.59	14/20=0.70	13/18=0.73	11/18=0.62	19/20=0.95*
		7	1:3/157=0.85	20/20=1.00	11/20=0.55**	10/20=0.50**	14/20=0.70**	20/20=1.00
		Q	133/160=0.84	18/20=0.90	13/19=0.69	17/20=0.85	14/20=0.70	16/20=0.80

SYMBOLS IN FIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HIST-FIGNI CONTEGL GROUP

ONE !,* > DIGNIFICANT AT P LESS THAN 0.05 TWO !,* > DIGNIFICANT AT P LESS THAN 0.01

^{*} SIGNICICATIVE DIFFERENT FROM CONTROL

[!] SIGNITION LINEAR RELATIONSHIP WITH ARITH OR LOG DOSE (HEADING OF COLUMN)

COMPOUND 40 STUDY ACUTE

AVERAGE NUMBER OF IMPLANTATIONS PER PREGNANT FEMALE.

	ARITH DOSE	h []	HISTORICAL CONTECL	NEGATIVE CONTROL		OSE LEVEL DO 500.000 MG/KG 50	DOO.000 MG/KG	POSITI VI CONTPOL
<u>!</u> !	?	1	1351/109=12.4	171/13=13.2	134/12=11.2	159/13=12.2	96/ 9=10.7an	52/10= 5.2**c **;
		2	1427/119=12.0) 183/16=11.4	176/15=11.7	203/16=12.7	155/13=11.9	96/11= 7.0**D **3
; <u>!</u>		3	1435/119=12.1	1 217/16=13.6	148/12=12.3	165/14=11.8*მმნ) 186/15=12.4@D	93/11= 8.5*** ***()
, !	!	Ċ.	1626/136=12.0) 222/18=12.3	182/18=10.1*00D *a0D		181/17=10.7*aD aD	171/16=10.7*** ***
		5	1966/127=11.5	5 221/19=11.6	228/19=12.0	236/20=11.8	227/19=12.0	205/18=11.4
		÷	15 12/128-11.8	3 138/13=10.6	178/14=12.7*@I *@I	156/13=12.0	126/11=11.5	167/19= 8.800 ***
• .		7	1626/133=12.2	2 252/20=12.6	133/11=12.1	113/10=11.3@D *@D	167/14=11.9	205/20=10。3#38 **7
		3	1551/133=11.7	7 213/18=11.8	157/13=12.1	198/17=11.7	160/14=11.4	205/16=12.8

SYMBOLS OF FIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS OF SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HIST HIGH CONTROL GROUP

S AND * = TWO-TAILED TEST ! AND @ = ONE-TAILED TEST

 $\stackrel{\bullet}{=}$ 1.8, ,* = SIGNIFICANT AT P LESS THAN 0.05 TWO 1.8, ,* = SIGNIFICANT AT P LESS THAN 0.01

. , & SIG' | IC'' TLY DIFFEIENT FROM CONTROL

o, i sig . . . n Pelonichagin wimm spina holdes deen (Beartya he coigra)

COMPOSITE 46 STITE ACUTE

AVERAGE CORPORA LUTEA PER PREGNANT FINALE

LOG AFITH DOSE DOSE		HISTORICAL CONTROL	NEGATIVE CONTPOL		POSE LEVEL : 1 500.000 MG/FG 5	OOSE LEVEL 5000.000 MG/KG	POSITIVE CONTROL
	1	1504/109=13.8	173/13=13.3	168/12=14.0	184/13=14.2	121/ 9=13.4	31/10= 9.1*** **
!	2	1588/119=13.3	202/16=12.6	185/15=12.3	204/16=12.8	160/13=12.3 ap	129/11=11.7 * o i
!	2	1365/119=13.2	217/16=13.6	151/12=12.6	169/14=12.1*35 #DI	•	116/11=10.6*2r *2P
8 ! 88!! 88!!	Ŀ	1784/ 13 6= 13.1	225/18=12.5	199/18=11.0*მმ **ა		18 7/17=11.0* 000 000 **00	
	5	1048/127=13.0	230/19=12.1	234/19=12.3	247/20=12.4	234/19=12.3	209/18=11.6 ******
1 1136	6	1089/12d=13.2	180/13=13.9	221/14=15.8 **@	194/13±14.9 ∂I ∂I	166/11=15.1 *@@I	213/19=11.2†35 ***
	7	1767/133=13.3	255/20=12.8	156/11=14.2&T	151/10≃15.1*∂] *∂]		250/20=12.5
38!!	ರೆ	1823/133=13.7	319/18=17.7		DD 242 /17=14. 2*∂!	203/14=14.5*@D	243/16=15.2

SYMBOLS OF LIPST LINE DENOTE SIGNIFICANT PELATIONSHIPS AND DIFFERENCES USING THE NEGITIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DEMOTE SIGNIFICANT PELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

```
S AND * = TWO-TAILED TEST
! AND * = OWE-TAILED TEST
```

CUP 1,8,., = SIGNIFICANT AT P LESS THAN 0.05 TWO 1,8, . = SIGNIFICANT AT P LESS THAN 0.01

*, D SIGNARY THE DIFFERENT FROM CONTROL

S.! SITE OF THE PRINCIPATION WITH ADDRESS DOOR (PEARLING OF COLUMN)

AVERAGE PREIMPLANTATION LOSSES PER PREGNANT FEMAL!

		HISTOPICAL CONTROL	NEGATIVE :	OSE LEVE 50.000	L DOS	SF LEVF!	DOSE MG/FG 5000	.000 MG		CONTRO!	
8 ! !	1	153/109= 1.4	2/13= 0.2 ***		2.8**@DI	25/13=	1.9**@NI	25/ 9=	2.8**@%I	39/10=	3. ○★★?) ★☆☆~
ε!!	2	161/119= 1.4	19/16= 1.2	3/15=	0.6 *ap	1/16=	0.1 **@@D	5/13=	ი. u **შეე	43/11=	3.9★☆ [™] *?T
811 8 1	3	130/119= 1.1	0/16= 0.0	3/12=	0.3 **aaD	4/14=	0.3 **⊋⊕D	4/15=	C.3 ************************************	23/11=	2.1*200
ε!! ε !	34	158/ 13 6= 1. 2	3/18= 0.2 **@dI	16/18=	0.9	0/19=	0.0 **aaD	6/17=	0.4 **30D	9/16=	0.6
8!: !	5	182/127= 1.4	9/19= 0.5 **∂ðI	6/19=	0.3 **aap	11/20=	0.6 *@@D	7/19=	0.4 **@@!	4/18=	0.2 ***
811 88 11	6	177/128= 1.4	42/13= 3.2 *9I	43/14=	3.1 **@nI	38/13=	2.° **∂∂I	40/11=	3.6 **aaT	46/19=	2.4
8!! 8!!	7	141/133= 1.1	3/20= 0.2 **aa	23/11=	2.1**00I **00I	38/10=	3.8**00I **00I	18/14=	1.3*aaI	45/20=	2.3************************************
i	٥	272/133= 2.1	106/18= 5.9 **@@	19/13= I	1.5**@3D	44/17=	2.6*aD	43/14=	3.12D	38/16=	2. 4* 图8

SYMBOLS ON TIPST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISPORICAL CONTROL GROUP

8 AND * = TYO-TAILED TEST ! AND B = ONE-TAILED TEST

ONE 1.5. , $^{\circ}$ = SIGNIFICANT AT P LESS THAN 0.05 TWO 1.8. , $^{\circ}$ = SIGNIFICANT AT P LESS THAN 0.01

THE SIGN TIPES THE DIFFERENT FROM CONTROL OF LOG DOWN (FINDING OF COLUMN)

orrowro 40 smary Achre

AVERAGE RESORPTIONS (DEAD IMPLANTS) PER PRESNANT FEMALE

	ARITH DOSE	X 777	HISTOPICAL CONTROL	NEGATIVE CONTECL	DOSE LEVEL 50.000 MG/KG	200.000 MGNKG DOSE TEADT	DOSE FIAST POOC*OCC WC/KC	CONTROL BOSITIVE
1 !		7	28/109=0.26	3/13=0.24	5/12=0.42	11/13=0.85*3I *@@I	3/ 9=0.34	42/10=4.20**0@I **@@T
	8 !! 8 !	2.	53/119=0.45	4/16=0.25	4/15=0.27	3/16=0.19 @D	17/13=1.31	36/11=3.28**37I **?7I
		2	(1/119±0.52	10/16=0.63	13/12=1.09	2/14=0.15 *33D	15/15-1.00	14/11=1.28 ****
8 ! ! 8 ! !	ε!!		62/136=0.46	12/18=0.67	5/18=0.23	େ/19=0.0 *⊅ର **∂ଚା	1/17=0.06*aD **aaD	73/16=4.57**@@I **@@I
811	8 !	۲,	74 /127 =0.59	6/19=0.32	3/19=0.16 **aa	7/20=0.35	2/19=0.11 **@@D	16/18=0.89
!		FX	53/128=0.46	2/13=0.16 *aD	4/14=0.29	3/13=0.24	2/11=0.19	23/19=1.23**90I **90I
!		7	A5/133=0.49	1/20=0.05 **@@	3/11=0.28	6/10=0.60	8/14=0.580I	1/20=0.05 **33D
		~ <u>`</u>	71/133=0.54	5/18=0.28	5/13=0.39	10/17=0.59	5/14=0.36	13/16=0.82

SYMBOLS OF THESE TIME DENOTE SIGNIFICANT PELATIONSHIPS AND DIFFERENCES USING THE NEGRETIC CONTROL GROUP

SYMBOLS OF SECOND LINE DENOTE SIGNIFICANT FELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTECL GROUP

S AND * = TVO-TAILED TEST ! AND D : OUF-TAILED TEST

THE 1.8, . = SIGNIFICANT AT PLESS THAN 0.05 THO 1.8, . = SIGNIFICANT AT PLESS THAN 0.01

-, & SIG (100 TIM DIVUENENT PROM CONTROL D.! SIG () (BEADING OF COLUMN)

δ

PROPORTION OF FEMALES WITH CNF OF MORE DEAD IMPLANTATIONS

	ARITH DOSE W	€ इंद	PISTOPICAL CONTROL	NEGATIVE CONTROL	DOSE LEVEL 50.000 MG/KG	DOSE LEVEL 500.000 MG/KG	POSE LEVEL	POSITIVE CONTROL
		1	24/109=0.23	3/13=0.24	3/12=0.25	8/13=0.62*	3/ 9=0.34	8/10=0.80**
		Ž	33/119=0.32	3/16=0.19	4/15=0.27	3/16=0.19	6/13=0.47	7/11=0.64*
		3	39/119=0.33	5/16=0.32	2/12=0.17	2/14=0.15	3/15=0.20	7/11=0.64
: !		4	46/136=0.34	6/18=0.34	5/18=0.28	0/19=0.0 **	1/17=0.06*	14/15=0.88** **
! !	! !	Ç,	45/12 7 =0.36	4/19=0.22	2/19=0.11	4/20=0.20	2/19=0.11	8/18=0.45
		G	44/128=0.35	2/13=0.16	3/14=0.22	3/13=0.24	2/11=0.19	13/19=0.69**
		7	46/133=0.35	1/20=0.05	3/11=0.28	1/10=0.10	4/14=0.29	1/20=0.05
		ų	50/133=0.38	4/18=0.23	4/13=0.31	7/17=0.42	5/14=0.36	6/16=0.38

SYMBOLS ON FIRST LINE DENOTE SIGNIFICANT PELATIONSHIPS AND DIFFERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HIST FIGAL CONTECT GROUP

ONE !, # - SIGNIFICANT AT P LESS THAN 0.05 TWO !, # - SIGNIFICANT AT P LESS THAN 0.01

^{*} SIGNI TIMELY DIFFFRENT FROM CONTROL

[!] SIGNI, CAN'T LIMEAR PELATIONSHIP WITH ARITH OF LOG DOSE (HEADING OF COLUMN)

COMPOUNT 40 COMPUT TO STUDY DOWN F

PORPORTION OF FEMALES WITH INO OR MORE DEED IMPLANTATIONS

APITH DOSE USES	BISTORICAL CONTROL	CONTROL	DODE LEVEL 50.000 MG/KG	• • • • • • • • • • • • • • • • • • • •	DOGE LEVEL	COMIDOR
1	3/109=0.03	0/13=0.0	2/12=0.17	2/13=0.16	0/9=0.0	7/10=0.70**
2	14/119=0.12	1/16=0.07	0/15=0.0	0/16=0.0	2/13=0.16	7/11=0.64**
3	17/119=0.15	4/16=0.25	1/12=0.09	0/14-0.0 *	1/15=0.07	5/11=0.45
<u>s</u>	12/136=0.09	3/18=0.17	0/18=0.0	0/19=0.0	0/17=0.0	13/16=0.82**
ϵ_{j}	18/127±0.15	2/19=0.11	1/19=0.06	1/20=0.05	0/19=0.0	4/18=0.23
44	13/128=0.11	0/13=0.0	1/14=0.08	0/13=0.0	0/11=0.0	5/19=0.32* **
7	14/133=0.11	0/20=0.0	0/11=0.0	1/10=0.10	2/14=0.15	0/20=0.0
9	18/133±0.14	1/18=0.06	1/13=0.08	2/17=0.12	0/14=0.0	4/16=0.25

SYMBOLS OF BILDY LINE DINOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE NPGLOLYF CONTROL GROUP

SYMBOLS ON RECORD LINE PENOTE SIGNIFICANT BELATIONSHIPS AND DIFFERENCES USING THE HISCOTTECL CONTROL GROUP

ONE !.* A DISCIPLICANT AT PLESS THAN 0.05 TAO !.* - DISCIPLICANT AT PLESS THAN 0.01

^{*} SIGNIC CONTROL DIFFERENT FROM CONTROL

[!] SIGNITED TO LINEAR RELATIONSPIP WITH ARITH OF LOG DOSE (BEADING OF COLUMN)

DARDOUND 40 TIUDY ACRES

DEAD IMPLANTS / TOTAL IMPLANTS

6 - TF	HISTOPICAL CONTECL	NEGATIVE CONTECL	DOSE LEVEL 50.000 MG/KG	DOSE LEVEL 500.000 MG/KG	DOSF LEVEL 5000.000 MG/KG	COMIBOT BOSIMIAL
1	28/1351=0.03	3/171=0.02	5/134=0.04	11/159=0.07*@ *@		42/ 52=0.81** **?
2	53/1427=0.04	4/183=0.03	4/176=0.03	•	17/155=0.11 @aD	36/ 86=0.42** **?
3	61/1435=0.05	10/217=0.05	13/148=0.09	•	15/186=0.09	14/ 93=0.16#* ***J
4	62/1626=0.04	12/222=0.06	5/182=0.03	0/222=0.0 @n **	1/181=0.01 @@D **@@!	73/171=0.43** ***
5	74/1466=0.06	6/221=0.03 *@D	3/228=0.02	7/235=0.03	2/227=0.01 **@@I	16/205=0.08 <i>ā</i> ī
f.	58/ 151 2=0.04	2/138=0.02	4/178±0.03 @D	3/156=0.02 *∂	2/126=0.02	23/167=0.14** ***
7	65/1626=0.04	1/252=0.01 **a@		6/113=0.06	8/167=0.05@I	1/205=0.016T **/
8	71/1551=0.05	5/213=0.03	5/157=0.04	10/198=0.06	5/160=0.04	13/205=0.07

SYMBOLS ON FIRST LINE DENOTE SIGNIFICANT DIFFERENCES USING THE NEGOTIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT DIFFERENCES USING THE HIST FIGURE CONTECL GROUP

^{# =} TWO+TALLED TEST
= ONF-LALLED TEST

O'E *, Φ = GIGNIFICANT AT P LESS THAN 0.05 TWO *, Φ + DIGNIFICANT AT P LESS THAN 0.01

^{*,} D SIG TATIONTY DIFFERENT FROM CONTROL

TODER ALLE TO SOUDA RABVOOLE

FERTILITY INDEX

OG APITH OSE MOSE WHER	HISTOPICAL CONTROL	NEGATIVE CONTROL	DOSE LEVEL 50.000 MG/KG	DOSE TEVEL 500.000 MG/KG	
1	1-4/159=0.66	13/19=0.69	12/19=0.64	8/19=0.43	11/20=0.55
2	118/160=0.74	14/20=0.70	18/20=0.90	13/20=0.65	14/20=0.70
3	119/159=0.75	12/19=0.64	19/20=0.90*	16/20=0.90	19/20=0.95* *
4	120/154=0.78	16/19=0.85	16/20=0.80	14/19=0.74	19/20=0.95
5	122/157=0.78	18/20=0.90	19/20=0.95	15/19=0.79	19/20=0.95
•,	134/159=0.86	18/19=0.95	14/20=0.70*	16/20=0.80	20/20=1.00
! 7 !	1°5/155=0.88	19/19=1.00	13/20=0.65**	12/20=0.60**	13/20=0.65**

SYMBOLS ON FIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE NEGRITUD CONTROL GROUP

SYMBOLS OF STOOMS LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFFRENCES USING THE HISTOTICAL CONTROL GROUP

CTE !,* = SIGNIFICANT AT P LESS THAN 0.05 TWO !,* = SIGNIFICANT AT P LESS THAN 0.01

^{*} SIGNICULATINY DIFFERENT FROM CONTROL

[!] SIGNIE COUNT LINEAR PELATIONSHIP WITH ARITH OR LOG DOSF (HEADING OF COLUMN)

TABLE TI COMPOUND 40 STUDY SUBJUSTED

AVERAGE NUMBER OF IMPLANTATIONS PER PREGNANT FUMALE.

	ARITH		HISTOFICAL CONTROL	NEGATIVE CONTECL		DOSE LEVEL 500.000 MG/KG	DOSE LEVEL 5000.000 MG/KG
	ε!	1	1231/104=11.8	167/13=12.9	160/12=13.3 **@	105/ 8=13.1	
		2	1474/118=12.5	190/14=13.6 @I	234/18=13.0	159/13=12.207	178/14=12.7
		3	1405/119=11.8	139/12=11.6	225/18=12.5	189/16=11.8	226/19=11.9
		4	1414/120=11.8	191/16=11.9	193/16=12.1	177/14=12.6	232/19=12.2
		5	1462/122=12.0	219/18=12.2	241/19=12.7	195/15=13.0 *a	233/19=12.3
; !	!	6	15.25/136=12.0	179/19= 9.9 **@	135/14= 9.6 0D **a		230/20=11.591
!	5 !	7	1566/135=11.6	204/19=10.7 an	144/13=11.1	144/12=12.001	130/13=10.0 ap

SYMBOLS ON FILST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

- S AND * = TWO-TAILED TEST ! AND D = OUM-TAILED TEST
- OUF !.S. * = SIGNIFICANT AT P LFSS THAN 0.05 TWO !.S. .. * = SIGNIFICANT AT P LESS THAN 0.01
- *, D SIGNIMICANTLY DIFFFERNT FROM CONTROL

 8,! SIGNIMICANT PELATIONSHIP WITH ABITH OR LOG DOSE (HEADING OF COLUMN)

COMPOUND 40 STUDY SUBACUIT

AVERAGE COPPORA LUTTA PER PREGUANT REVALE

	/RITH	W TE K	HISTORICAL CONTROL	NEGATIVE CONTROL		TOSE LEVEL 500.000 MG/KG	
	88!! 8 !	. 1	1385/104=13.3	171/13=13.2	167/12=13.9	112/ 8=14.0	126/11=11.50D **00D
		2	1599/118:13.6	194/14=13.9	234/13=13.0	166/13=12.8	180/14=12.9
		3	1935/119=12.9	139/12=11.6 *@@D	227/18=12.6	196/16=12.3	233/19=12.3
		ŭ	1499/120=12.5	198/16=12.4	198/16=12.4	177/14=12.6	237/19=12.5
:		5	1554/122=12.7	219/18=12.2	248/19=13.10I	195/15=13.0	250/19=13.2@I
; !!		ě,	1:09/136=13.3	193/18=10.7 **@3		200/16=12.5*@	I 257/20=12.9**aaI
!		7	1711/135=12.7	215/19=11.3 *ab	154/13=11.9		00I158/13=12.2 ∂@I

SYMBOLS OF FIRST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS AN SECOND LINE DENOTE SIGNIFICANT PELATIONSHIPS AND DIFFERENCES USING THE HISTORICAL CONTROL GROUP

8 AND * = TWO-TAILED TFST
! AND @ = OUF-TAILED TEST

ONE 1.8, . * = SIGNIFICANT AT P LESS THAN 0.05 THO 1.8, . * = SIGNIFICANT AT P LESS THAN 0.01

*,@ SIG INDIANTLY DIFFERENT FROM CONTROL

8,! SIGNIFICANT PELATIONSHIP WITH APITH OF LCG DOSF (HEADING OF COLUMN)

COUROUND 40

STUDY SUBACUTE

AVERAGE PREIMPIANTATION LOSSES PER PREGNANT FEMALE

			HISTORICAL CONTROL		ST LEVEL DOS 50.000 MG/KG 50	SE LEVEL DOSE 06.000 MG/KG 5000	0.000 MG\ke HEAST
:8!!	25!!	1	154/104= 1.5	4/13= 0.3	7/12= 0.6 *3D	7/8= 0.9	೧/11= 0.0⊅D **∂∂ŋ
38!!	!	2	125/116= 1.1	4/14= 0.3 **ສ໖ກ	0/18= 0.0 **@@D	7/13= 0.5	2/14= 0.1 **∂∂D
38!!	s !	?	130/119= 1.1	0/12= 0.0 **@@D	2/18= 0.1 **@@n	7/16= 0.4 *@D	7/19= 0.4 **aab
; !!		4	85/120= 0.7	7/16= 0.4	5/16= 0.3	0/14= 0.0 **@3D	5/19= 0.3 mp
!	E !	5	92/122= 0.8	0/18= 0.0 **@@D	7/19= 0.4	0/15= 0.0 **@@D	17/19= 0.90I
·		6	183/ 13 6= 1.4	14/18= 0.8	38/14= 2.7*DI	21/16= 1.3	27/20= 1.4
;81: ;81:		7	145/135= 1.1	11/19= 0.6	10/13= 0.8	25/12= 2.1*ลิปิโ อิโ	28/13= 2.2*@@I *@I

SYMBOLS OF TIEST LINE DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE NEGATIVE CONTROL GROUP

SYMBOLS ON SECOND LINE DENOTE SIGNIFICANT PELATIONSHIPS AND PIFFEPENCES USING THE HISTORICAL CONTROL GROUP

8 AND * = TROTTAILED TEST : AND @ = ONE-TAILED TEST

ONE 1,8,0,* = SIGNIFICANT AT P LESS THAN 0.05 THO 1,8,1,* = SIGNIFICANT AT P LESS THAN 0.01

*, a SIGNATION DIFFERENT FROM CONTROL

S,! SIG INTOLUT RELATIONSHIP WITH ARITH OF LOG DOSE (HEADING OF COLUMN)

TAN TI - COMPOUNT - 40 SIUDY SUBNOUTE

PROPORTION OF FEMALES WITH ONE OR NOWE DEAD IMPLANTATIONS

LOG ARITH		HISTOLICAL CONTROL	NEGATIVE CONTECL	DOST LEVEL 50.000 MG/KG	DOSE LEVEL 500.000 MG/KG	DOST LEVEL 5000.000 MG/KG
	1	² 1/104=0.30	7/13=0.54	3/12=0.25	3/ 8=0.38	1/11=0.10*
	2	19/118=0.33	7/14=0.50	8/18=0.45	6/13=0.47	2/14=0.15*
! !	3	-2/ 119 =0.36	2/12=0.17	9/18=0.50	7/16=0.44	11/19=0.58*
	4	82/ 120= 0.35	6/16=0.38	4/16=0.25	0/14=0.0 *	7/19=0.37
!! !!	5	54/122=0.45	5/18=0.28	3/19=0.16	2/15=0.14	3/19=0.16 *
! !	4	43/136=0.32	1/18=0.06	1/14=0.08	0/16=0.0	4/20=0.20
	7	42/135=0.32	3/19=0.16	4/13=0.31	1/12=0.09	3/13=0.24

SYMBOLS IN CIRST LITE DENOTE SIGNIFICANT PELATIONSHIPS AND DIFFERENCES USING THE NEGRTIVE CONTROL GROUP

SYMBOLS ON SECOND LIME DENOTE SIGNIFICANT RELATIONSHIPS AND DIFFERENCES USING THE HISTORIUM CONTROL GROUP

ONE !.* = SIGNIFICANT AT P LESS THAN 0.05 TWO !.* = CIGNIFICANT AT P LESS THAN 0.01

^{*} SIGNIFICATORY DIFFERENT FROM CONTROL

[!] SIGNIFICANT LINEAR RELATIONSHIP WITH ABITH OF LOG DOSE (HEADING OF COLUMN)

TARIF VII

COMPOUND 40 STUDY SUPLCUTE

PORPORTION OF FEMALES WITH TWO OF MOSE DEAD IMPLANTATIONS

LOG ARI		HISTODICAL CONTROL	NEGATIVE CONTECL	DOSE LEVEL 50.000 MG/KG	DOSE LEVEL 500.000 MG/KG	DOSE LEVEL 5000.000 EG/KG
	1	8/104=0.08	1/13=0.08	1/12=0.09	1/8=0.13	0/11=0.0
	2	10/118=0.09	4/14=0.29	6/18=0.34	1/13=0.08	1/14=0.08
!!	. 3	17/119=0.15	1/12=0.09	7/18=0.39	2/16=0.13	8/19=0.43*
	<u></u>	15/120=0.13	1/16=0.07	0/16=0.0	0/14=0.0	3/19=0.16
	5	19/122=0.16	2/18=0.12	2/19=0.11	1/15=0.07	1/19=0.06
	5	13/136=0.10	0/18=0.0	1/14=0.08	0/16=0.0	0/20=0.0
	7	16/135=0.12	2/19=0.11	1/13=0.08	0/12=0.0	0/13=0.0

SYMBOLS ON FIRST LINE DENOTE SIGNIFICANT PELATIONSHIPS AND DIFFERENCES USING THE NEG TIVE COUTROL GROUP

SYMBOLS OF SECOND LINE DENOTE SIGNIFICANT PELATIONSHIPS AND DIFFERENCES USING THE HIS GOICLE CONTROL GROUP

CNE 1.* = DIGNIFICANT AT P LESS THAN 0.05 TWO 1.* = DIGNIFICANT AT P LESS THAN 0.01

^{*} SIGNI ICANTLY DIFFERENT FFOR CONTROL

[!] SIGNE TOTAL LINEAR RELATIONSHIP WITH ARITH OF LOG DOSE (HEADING OF COLUMN)

DEAD IMPLANTS / TOTAL IMPLANTS

মুণু ত≱	HISTOPICAL CONTRUL	NEGATIVE I	DOSE LEVEL 50.000 MG/KG	DOSE LEVEL 500.000 MG/KG	DOSE MEVEL - ROCO.OCO MG/KG
1	40/1231=0.04	8/167=0.05	4/160=0.03	4/105=0.04	1/126=0.01&D *&D
2	59 /1474 =0.05	18/190=0.10	31/234=0.14	7/159=0.05	3/178=0.02@D ap
7	69/1405=0.05	3/139=0.03	70/225=0.32*@@ *@@		60/226=0.27*∂@I *∂I
f : ∳	r6/1414=0.05	7/191=0.04	4/193=0.03		:*@@D11/232=0.05 :*@@D
5	7//1462=0.06	7/219=0.04 ap	5/241=0.03 **d		4/233=0.02 **ଅଟମ *ଅଟମ
€.	(2/1626±0.04	1/179=0.01 *@@D	4/135=0.03	· ·	4/230=0.02 **@DD
7	70/1566=0.05	5/204=0.03	5/144=0.04	1/144=0.01	3/130=0.03 *≈∂∂D

SYMBOLS OR FIRST LINE DENOTE SIGNIFICANT DIFFERENCES USING THE NEGROUP CONTROL GROUP

SYMBOLS OF GROUP LINE DENOTE SIGNIFICANT DIFFERENCES USING THE HISTORICAL CONTECT GROUP

* = TWO+.AILLO TEST B = ONE-TAILLE TEST

OTE *, 0 = DISCIPICANT AT P LESS THAN 0.05 TWO *, 0 : DISCIPICANT AT P LESS THAN 0.01

*. D SIGHTFICHHTLY DIFFERENT FROM CONTROL

II. APPENDICES (MATERIALS AND METHODS)

A. Animal Husbandry

1. Animals -- Rats and Mice

Ten to twelve week old rats (280 to 350 g) and male mice (25-30 g) were fed a commercial 4% fat diet and water ad libitum until they were on experiment. Flow Laboratories random-bred, closed colony, Sprague-Dawley CD strain rats were used in the cytogenetic studies. Flow Laboratories ICR male mice were employed in the Host-Mediated Assay.

2. Preparation of Diet

A commercial 4% fat diet was fed to all animals. Periodic tests to verify the absence of coliforms, <u>Salmonella</u> and <u>Pseudomonas</u> sp. were performed.

3. Husbandry

Animals were held in quarantine for 4-11 days. Mice were housed five to a cage and rats one to five to a cage. Animals were identified by ear punch. Sanitary cages and bedding were used, and changed two times per week, at which time water containers were cleaned, sanitized and filled. Once a week, cages were repositioned on racks; racks were repositioned within rooms monthly. Personnel handling animals or working with animal facilities wore head covering and face masks, as well as suitable garments. Individuals with respiratory or other overt infections were excluded from the animal facilities.

B. Dosage Determination

1. Acute LD_{50} and LD_{5} Determination Since the compounds proposed for testing are included in the



food additive regulations as "generally recognized as safe" (GRAS), it was expected that a large number of them would be sufficiently non-toxic so that determination of an LD_{50} or an LD_{5} is of no practical value. In fact, this has been our experience with previously tested compounds from this list. In the case of these relatively non-toxic compounds, attempts were made to assure that the amounts to be administered would not affect the animals by means (mechanical, physical, etc.) related to their bulk rather than to their toxicity. In the cases of certain compounds where an LD_{50} or an LD_{5} were not determined, an exceedingly high concentration, 5 g/kg, was employed and accepted as the LD_{5} level. In cases where the toxicity was high enough to allow determination of an LD_{50} , the following protocol was used.

Thirty rats of the strain chosen for studies described below and of approximately the age and weight specified were assigned at random to six groups. Each group was then given, using the chosen route of administration, one of a series of dosages of the test compound following a logarithmic dosage scheme. The series of dosages was derived from a consideration of whatever toxicity information was available for the particular test compound. The objective in selecting dosages was to choose values which would cause mortalities between 10% and 90%.

When information was inadequate to derive a suitable series of dosages, five rats were used to identify the proper range. Each of these was given one of a widely spaced (differing by 10X) series of doses. This was confidently expected to suffice for derivation of the series of dosages to be used in the LD_{50} determination.

The mortalities observed when the series of dosages was given to the 30 rats were then subjected to a probit analysis and calculation of LD_{50} , LD_{5} , slope and confidence limits by the method of Litchfield and



Wilcoxon. The highest dose level used was either a finite LD $_5$ or 5000mg/kg. The intermediate level used was either 1/10 of the finite LD $_5$ or 500mg/kg. The low level used was either 1/100 of the finite LD $_5$ or 50 mg/kg.

2. Subacute Studies

Subacute doses were identical to those used in the acute studies. Each subacute study animal was given the acute dosage once a day for each of five consecutive days (24 hours apart).

C. <u>Mutagenicity Testing Protocols</u>

1. Host Mediated Assav

Flow Laboratories ICR random-bred male mice were used in this study. In the acute and subacute studies ten animals, 25-30 g each, were employed at each dose level. Solvent and positive controls were run at all times. The positive control (Dimethyl nitrosamine) was run by the acute system only at a dose of 100 mg/kg for Salmonella. For yeast, ethyl methane sulfonate (EMS) intramuscularly injected at a dose of 350 mg/kg was used. The solvents used and the toxicity data are presented in the Results and Discussion section of the report.

The indicator organisms used in this study were: (1) two histidine auxotrophs [his G-46, TA-1530] of Salmonella typhimurium, and (2) a diploid strain [D-3] of Saccharomyces cerevisiae. The induction of reverse mutation was determined with the Salmonella; mitotic recombination was determined with yeast. Chemicals were evaluated directly by in vitro bacterial and yeast studies prior to, or concurrent with, the studies in mice. Animals on acute studies only were not fed the evening prior to compound administration. The Salmonella were carried in tryptone yeast extract gel, transferred weekly. They were transferred to tryptone yeast extract broth 48 hours



prior to use, and again 8 hours before use. The mouse inoculum was prepared by transferring 4 ml of the 8-hour broth culture to 50 ml broth bottles which had been prewarmed to 37°C. Exponential log-phase organisms were inoculated intraperitoneally into the mice approximately 2-1/2 hours later when the appropriate density indicating 3.0 x 10⁸ cells/ml was reached. The <u>Saccharomyces</u> was carried in yeast complete agar. The inoculum was prepared by harvesting the organisms from the surface of the plates with sterile saline. The cells were washed three times with sterile saline and suspended in a concentration of 5.0 x 10⁸ cells/ml. Two ml of the suspension was inoculated into each mouse intraperitoneally. Total plate counts on <u>Salmonella</u> were on tryptone yeast extract and for <u>Saccharomyces</u> on yeast complete medium.

a. Acute Study

Three dosage levels (usage, intermediate [determined as discussed previously], and LD $_5$) were administered orally by intubation to ten mice. Positive controls and negative vehicle controls were included in each study. All animals received 2 ml of the indicator organism intraperitoneally. Each ml contained 3.0 x 10^8 cells for Salmonella and 5.0 x 10^6 cells for Saccharomyces. Three hours later, each animal was killed and 2 ml of sterile saline was introduced intraperitoneally. As much fluid as possible was then aseptically removed from the peritoneal cavity. Dilution blanks for bacteria containing 4.5 ml of sterile saline were prepared in advance. Tenfold serial dilutions were made of each peritoneal exudate (0.5 ml exudate + 4.5 ml saline)



yielding a concentration series from 10^{0} (undiluted peritoneal exudate) through 10^{-7} . For enumeration of total bacterial counts, the 10^{-6} and 10⁻⁷ dilutions were plated on tryptone yeast extract agar, 3 plates/sample, 0.2 ml sample/plate. Each sample was spread over the surface of the plate using a bent glass rod immersed in 95% ethanol and flamed just prior to use. In plating for the total mutant counts on minimal agar, the 10^{0} dilution was used, 0.2 ml being plated on each of 5 plates. The plating procedure was identical to that followed for the tryptone yeast extract agar plates. All plates were incubated at 37°C, tryptone yeast extract agar plates for 18 hours and minimal agar plates for 40 hours. For yeast mitotic recombination, dilution blanks containing 4.5 ml of sterile saline were prepared in advance. Tenfold serial dilutions were made of each sample yielding a series from 10^0 to 10^{-5} . Samples of 0.1 ml of the 10^{-5} , 10^{-4} , and 10^{-3} dilutions were removed and plated on complete medium (10 plates each). All plates were incubated at 30° C for 40 hours. The 10^{-5} dilutions were used to determine total populations and the 10^{-4} and 10^{-3} plates were examined after an additional 40 hours at 4°C for red sectors indicating a mutation. Bacterial scoring was calculated as follows:

Total mutants on 5 plates X appropriate exponent = CFU/ml of sample plated (CFU is Colony Forming Units)

CFU/ml X one/dilution factor $(10^0 - 10^{-7})$ = CFU/ml in undiluted exudate. The mutation frequency (MF) was calculated for each sample where:



Yeast mitotic recombinants (presumptive <u>ade 2</u>, <u>his 8</u> homozygotes) were seen as red colonies or as red sectors on a normally white yeast colony. The plates (from 10^{-4} and 10^{-3} dilutions) were scanned under the 10X lens of a dissecting scope to enumerate the red colonies and sectors. Population determinations were made from the 10^{-5} dilution plates. A recombinant frequency (RF) was calculated:

RF = total recombinants counted total number colonies screened

b. Subacute Study

Similar groups of animals at each dose level received five oral doses of the test compound 24 hours apart. Within 30 minutes after the last dosing, the animals were inoculated with the test organism and handled in the same fashion as those in the acute study.

c. In Vitro Study

Cultures of <u>S</u>. <u>typhimurium</u> histidine auxotrophs (G-46 and TA-1530) were plated on appropriate media. The test compound was then added to the plate, either in the form of a microdrop of solution (0.01 to 0.25 ml) applied to a small filter paper disc resting on the agar or a small crystal applied directly to the agar. Tenfold serial dilutions of the culture were employed and plated so as not to miss the optimum cell density for mutant growth. Mutant colonies were observed and scored. Strain D-3 <u>Saccharomyces</u> cells at proper dilutions were shaken with the test compound, diluted, and plated at 50% survival level or above (see HMA Supplementary Materials and Methods). Red sectors were then scored and the frequency calculated after suitable incubation. Negative and positive controls were run concurrently. The positive control was EMS for <u>Salmonella</u> and <u>Saccharomyces</u>. The <u>in vitro Salmonella</u> tests were reported as (+) or (-) or questionable; the <u>in vitro Saccharomyces</u> tests were reported as sample concentrations, percent survival,



and recombinants/ 10^5 survivors. For the <u>Saccharomyces</u> a 50% survival level, e.g., an arbitrary 5.0% w/v test level, was used when no LD₅₀ was determinable.

2. Cytogenetic Studies

a. In Vivo Study

Ten to twelve week old, male, albino rats obtained from a closed colony (random-bred) were used. A total of 59 animals in the acute study and 18 animals in the subacute study was used, as illustrated in the following protocol.

Number of Animals Used

Acute Study

Treatment	Time Kille	ed after Admir	nistration
	6 hours	24 hours	48 hours
High level	5	5 ,	5
Intermediate level	5	5	5
Low level	5	5	5
Positive control	0	0	5
Negative control	3	3	3

Subacute Study

Five doses 24 hours apart; animals killed 6 hours after last dose.

Treatment	Killed after Administration
High level	5
Intermediate level	5
Low level	5
Negative control	3

All animals were dosed by gastric intubation.

Four hours after the last compound administration, and two hours prior to killing, each animal was given 4 mg/kg of colcemid intraperitoneally in



order to arrest the bone marrow cells in C-mitosis. Animals were killed by using CO₂, and the adhering muscle and epiphysis of one femur were removed. The marrow "plug" was removed with a tuberculin syringe and an 18 gauge needle, aspirated into 5 ml of Hanks' balanced salt solution (BSS) in a test tube and capped. The specimens were centrifuged at 1,500 RPM in a table-top centrifuge for 5 minutes, decanted, and 2 ml of hypotonic 0.5% KCl solution was added with gentle agitation to resuspend the cells. The specimens were then placed in a 37°C water bath for 20 minutes in order to swell the cells. Following centrifugation for 5 minutes at 1,500 RPM, the supernatant was decanted and 2 ml of fixative (3:1 absolute methanol: glacial acetic acid) was added. The cells were resuspended in the fixative with gentle agitation, capped, and placed at 4°C for 30 minutes. The specimens were again centrifuged, decanted, 2 ml of prepared fixative was added, and the cells were resuspended and placed at 4°C overnight.

The following day the specimens were again centrifuged, decanted and 0.3 - 0.6 ml of freshly prepared fixative was added to obtain a suitable density. The cells were resuspended and 2 - 3 drops of the suspension were allowed to drop onto a clean, dry slide held at 15° from the horizontal. As the suspension flowed to the edge of the slide, it was ignited by an alcohol burner and allowed to flame. Following ignition, the slides were allowed to dry at room temperature overnight. Duplicate slides were prepared. The slides were stained using a 5% Giemsa solution (Giemsa buffer pH 7.2) for 20 minutes, rinsed in acetone, 1:1 acetone:xylene, and placed in fresh xylene for 30 minutes. The slides were then mounted using permount (Fisher Scientific) and 24 X 50 mm coverglasses. The coverglasses were selected to be 0.17 mm ± 0.005 mm in thickness by use of a coverglasses mi rometer.



The preparations were examined using Leitz Ortholux I & II microscopes with brightfield optics and xenon light sources. These specimens were scanned with 10X and 24X objectives and suitable metaphase spreads that were countable were then examined critically using 40X, 63X or 100X oil immersion flat-field apochromatic objectives. Oculars were either 12X or 16X widefield periplanatics and the tube magnification either 1X or 1.25X. The filters used were either a didymium (BG20) or a Schott IL570 m_{μ} interference filter.

The chromosomes for each cell were counted and only diploid cells were analyzed. They were scored for chromatid gaps and breaks, chromosome gaps and breaks, reunions, cells with greater than ten aberrations, polyploidy, pulverization, and any other chromosomal aberrations which were observed. They were recorded on the currently used forms and expressed as percentages on the summary sheets. Fifty metaphase spreads were scored per animal. Mitotic indices were obtained by counting at least 500 cells and the ratio of the number of cells in mitosis/the number of cells observed was expressed as the mitotic index.

Positive controls in the acute study consisted of animals which had been given the known mutagen Triethylene Melamine (TEM) administered intraperitoneally at a level of 0.30 mg/kg. Negative controls on the acute and subacute studies consisted of the vehicle in which the compound was administered. The dosage levels, solvents and toxicity data are included in the Results and Discussion section of the report.

b. <u>In Vitro Study</u>

Human embryonic lung cultures (WI-38) which were negative for adventitious agents (viruses, mycoplasma) which may interfere were used. These cells were employed at passage level 19. The cells had been transferred



using 0.025% trypsin and planted in 32 oz. prescription bottles containing 40 ml of tissue culture medium. When growth was approximately 95% confluent the cells were removed from the glass using trypsin, centrifuged, and frozen in tissue culture medium containing dimethyl sulfoxide (DMSO). Cells were frozen in vials in the vapor phase of liquid nitrogen at a concentration of 2 x 10^6 cells/ml. When needed, the vials were removed from liquid nitrogen, quick-thawed in a 37°C water bath, washed free of DMSO, suspended in tissue culture medium (minimal essential medium [MEM] plus 1% glutamine, 200 units/ml of penicillin and 200 μ g/ml of streptomycin and 15% fetal calf serum) and planted in milk dilution bottles at a concentration of 5 x 10^5 cells/ml. The test compound was added at three dose levels using three bottles for each level, 24 hours after planting. The dose levels required a preliminary determination of a tissue culture toxicity. This was accomplished by adding logarithmic doses of the compound in saline to a series of tubes containing 5×10^5 cells/ml which were almost confluent. The cells were examined at 24, 48, and 72 hours. Any cytopathic effect (CPE) or inhibition of mitoses was scored as toxicity. Five more closely spaced dose levels were employed within the two logarithmic dosages, the higher of which showed toxicity and the lower no effect. The solvents used and the range finding data are presented in the toxicity data report under Results and Discussion. The dose level below the lowest toxic level was employed as the high level. Logarithmic dose levels were employed for the medium and low levels.

Cells were incubated at 37°C and examined twice daily to determine when an adequate number of mitoses were present. Cells were harvested by shaking when sufficient mitoses were observed, usually 24 - 48 hours after planting, centrifuged, and fixed in absolute methanol:glacial acetic acid (3:1) for 30 minutes.



The specimens were centrifuged, decanted, and suspended in acetic acid-orcein stain (2.0%) and a drop of suspension placed on a clean dry slide. Selected coverglasses 0.17 mm in thickness were placed on the suspension and the excess stain gently expressed from the slide. The coverglasses were sealed with clear nail polish and examined immediately.

The microscopes, objectives, oculars, filters and light sources were enumerated under the metaphase description. Positive controls used were TEM (at a concentration of 0.1 mcg/ml dissolved in saline) and negative controls which consisted of the vehicle in which the test compound was dissolved, which was 0.85% saline. Data were reported on forms currently used and expressed as percentages on the anaphase summary sheets.

3. Dominant Lethal Assay

In this test, male and female random bred rats from a closed colony were employed. These animals were 10-12 weeks old at the time of use. Ten male rats were assigned to each of 5 groups; 3 dose levels selected as described above, a positive control (triethylene melamine) (TEM) and a negative control (solvent only). The positive control was administered intraperitoneally. Administration of the test compound was orally by intubation in both the acute study (1 dose) and in the subacute study (1 dose per day for 5 days). Following treatment, the males were sequentially mated to 2 female per week for 8 weeks (7 weeks in the subacute study).



Two virgin female rats were housed with a male for 5 days (Monday through Friday). These two females were removed and housed in a cage until killed. The male was rested on Saturday and Sunday and two new females introduced to the cage on Monday. It has been our experience that conception has taken place in more than 90% of the females by Friday and that the two day rest is beneficial to the male as regards subsequent weekly matings. Females were killed using CO_2 at 14 days after separating from the male, and at necropsy the uterus was examined for deciduomata (early deaths), late fetal deaths and total implantations.

Sufficient animals were provided in our experimental design to accomodate for any reduction in the number of conceptions. Each male was mated with two females per week, and this provided for an adequate number of implantations per group per week (200 minimum) for negative controls, even if there was a four-fold reduction in fertility of implantations. Results were analyzed according to the statistical procedures described in Supplementary Materials and Methods. Corpora lutea, early fetal deaths, late fetal deaths and total implantations per uterine horn were recorded on the raw data sheets, which are submitted separately.



D. Supplementary Materials and Methods

- 1. Host Mediated Assay In Vitro and Formulae
 - a. Bacterial in vitro plate tests.

This method has been published by Ames: The Detection of Chemical Mutagens with Enteric Bacteria, in <u>Chemical Mutagens</u>;

<u>Principles and Methods for Their Detection</u>, Vol. 1, Chapter 9, pp. 267-282, A Hollaender, Editor, Plenum Press, New York (1971).

- b. In vitro for Mitotic Recombination.
- (1) Strain D-3 was grown to stationary phase on complete medium agar plates at 30° C. (3-4 days). Cells were rinsed from the plates and washed twice in saline and cell concentration determined spectro-photometrically. (A standard curve previously determined for colony forming units versus % transmittance at 545 mu was easily used).
- (2) Cells from the concentration suspension were diluted appropriately into 0.067 M Phosphate buffer pH 7.2 to provide 5×10^7 cells/ml in a total of 25 ml.
- (3) The test chemical was first tested for 4 hours at 30° C, with shaking, at concentrations which permitted determination of the 50% survival level. Then, if not included in the first experiment, the compound was tested again only at the 50% survival level. If 50% survival level could not be determined, the arbitrary test level of 5% w/v was used.
- (4) Following treatment, cells were diluted and plated on complete agar medium for determination of total population and red sectors. Total surviving population was conveniently measured on plates of 10^{-4} and 10^{-5} dilutions using 0.2 ml per plate (5 plates)



and sectors determined on plates of 10^{-3} and 10^{-4} dilutions using 0.2 ml per plate (5 plates). Plates are incubated for 2 days at 30°C followed by a holding period of 2 days at 4°C to promote color development with limited enlargement of the colonies. Red sectors are scored by systematically scanning the plates with a dissecting microscope at 10X magnification.

- (5) The frequency of red sectors can then be calculated and may be expressed conveniently as sectors per 10^5 survivors for comparison with untreated controls.
- (6) Ethyl Methane Sulfonate (EMS) was employed as the positive control in both in vitro systems.
- c. Minimal Medium (Bacteria):

Spizizen's Minimal Medium

4X Salt Solution:

 (NH_4) SO_4 8.0 gm K2HPO4 56.0 qm KH2PO4 24.0 qm Na Citrate 4.0 gm Mg SÖ_A 0.8 gm Biotin 0.004 gm

H₂O qs to 1 liter Sterilize by autoclaving (121°C/15 min.)

Medium:

4X Salt Solution : 250 ml

5.0% Glucose (sterile) :100 ml (If histidine is added at concen-

tration of 30 mg/liter, this 1.5% Bacto-agar (sterile) : 650 ml becomes a complete bacterial

medium.)



2. Cytogenetics <u>In Vitro</u> Preparation of Anaphase Chromosomes (from Nichols, 1970)

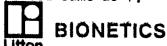
"Anaphase preparations may be made by several methods. One convenient approach is to grow cells directly on coverslips in petri dishes. With human fibroblasts 400,000 cells added to a 22 x 40 mm coverslip in a 50 mm petri dish grown in a 5% CO_2 atmosphere in air has proved very satisfactory. When adequate numbers of mitoses are visualized directly utilizing an inverted microscope (usually 48 to 92 hrs. after planting) the coverslip is transferred to absolute ethanol for 15 minutes for fixation. They are then stained with any one of a number of suitable stains (Fuelgen, May-Grunwald-Giemsa, orcein) and attached to a slide with mounting media for evaluation. Anaphase preparations may also be prepared on cells grown in suspension or cells from a monolayer that have been put into suspension. In this instance the cells are centrifuged and fixed with the squash fixative. They are then suspended in the stain and a drop of the suspension put on the slide and covered with a coverslip. However, in this case, only the excess stain is gently expressed from under the coverslip and no squashing is carried out. In anaphase preparations no pretreatment with colchicine or hypotonic expansion is used and no technique for spreading the cells is used, so that the spindle and normal relationships of the chromosomes are not disturbed."



3. Statistical Analyses of Dominant Lethal Studies

The following statistical analyses were employed as a means of analyzing the results of the dominant lethal studies.

- a. The fertility index: number of pregnant females/number of mated females with the chi-square test used to compare each treatment to the control. Armitage's trend used for linear proportions to test whether the fertility index was linearly related to arithmetic or log dose.
- b. Total number of implantations: t-test used to determine significant differences between average number of implantations per pregnant female for each treatment compared to the control. Regression techniques used to determine whether the average number of implantations per female was related to the arithmetic or log dose.
- c. Total number of corpora lutea: t-test used to determine significant differences between average number of corpora lutea per pregnant female for each treatment compared to the control.
- d. Preimplantation losses: computed for each female by subtracting number of implantations from number of corpora lutea. Freeman-Tukey transformation used on the preimplantation losses for each female and then t-test used to compare each treatment to control. Regression technique used to determine whether the average number of preimplantation losses per female was related to the arithmetic or log dose.
 - e. Dead implants: treated same as preimplantation losses.
- f. The proportion of females with one or more dead implants computed, each treatment compared to control by chi-square test and Armitage's trend used for linear proportions to see if proportions were linearly related to either arithmetic or log dose. Also, probit regression analysis used to determine whether the probit of the proportions was related to log dose.
- g. The proportion of females with two or more dead implants computed treated same as f.



h. Dead implants/total implants: computed for each female and used Freeman-Tukey arc-sine transformation on data for each female; then used t-test to compare each treatment to control.

Historical control data was compiled on a continuous basis as studies were completed. In addition to comparing each treatment to control, as outlined above, each treatment was compared to an historical control.

In order to take variation between males into account, a nested model was used. An analyses of across weeks is also provided.

In addition to these tests, the distribution forms of the various parameters were tested in order to evaluate the appropriateness of some of the tests being used. Certain correlations between parameters may exist and were examined as one step to determine the appropriateness of models. If necessary, alternate test methods were implemented.

The results are presented in tabular form with the addition of historical control information. In addition to these tables, a written report of all findings is provided. As information became available from the on-going investigation of these data, it was reported and suggestions included for changes to the methods of analyses. The statistical reports give the level of significance using both a one-tailed and two-tailed test. Finally, a summary sheet for each study is provided.



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L-1, 2 Groups

i -1, 2, ---, 10 Males within each group

K- 1, > Females within Males within Groups

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Males are randomly drawn from infinite population

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Abbreviations
mu = micron
mcg = ug = microgram
q = gram
kg = kilogram
ml = milliliter
rpm = revolutions per minute
OC = degrees centigrade
 pH = power of the hydrogen ion concentration to the base 10
 M = molar solution
 conc. = concentration
 MTD = maximum tolerated dosage = High = LD _{5} if determined or else exceedingly high dise, such as 5 g/kg
 INT = intermediate = medium level
 USE = usage level if known = low level
 BSS = balanced salt solution
 C-metaphase = cells arrested in metaphase, using colchine or colcemid
 LD<sub>50</sub> = that dosage which produced 50% mortality in the group of animals treated
 LD_5 = that dosage which produced 5% mortality in the group of animals treated
  NC = negative control
  PC = positive control
  AU = acute usage level (low level)
  AI - acute intermediate level (medium level)
  AMTD = acute maximum tolerated dose level (LD_5 level, high level)
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SAU = subacute usage level (low level) SAI = subacute intermediate level (medium level) $SALD_5$ = subacute LD_5 level (MTD level, high level) CO₂ = carbon dioxide DMN = Dimethyl nitrosamine EMS = Ethyl methane sulfonate TEM = Triethylene melamine DMSO = Dimethyl sulfoxide MEM = minimal essential medium (Eagle's) CPE = cytopathic effect his = histidine marker D-3 = mitotic recombinant strain of saccharomyces mf = mean mutant frequency MFt/MFc = mean mutant frequency of the test compound group compared to mean mutant frequency of the negative control group CFU = colony forming units WI-38 = code name for a strain of human embryonic lung tissue culture cells Rec \times 10⁵ = mitotic recombinants \times 10⁵ Mean B/A = mean frequency tot. scr. = total scored tot. = total = a test of variation in the data from the computed regression line. Tested in these studies at the 5% level Aberr. = aberrations Frag. = fragment HMA - host mediated assay

